



DHS SCIENCE AND TECHNOLOGY

Master Question List for COVID-19 (caused by SARS-CoV-2)

Weekly Report

21 April 2020

For comments or questions related to the contents of this document, please contact the DHS S&T Hazard Awareness & Characterization Technology Center at HACTechnologyCenter@hq.dhs.gov.



**Homeland
Security**

Science and Technology

DHS Science and Technology Directorate | MOBILIZING INNOVATION FOR A SECURE WORLD

CLEARED FOR PUBLIC RELEASE

FOREWORD

The Department of Homeland Security (DHS) is paying close attention to the evolving Coronavirus Infectious Disease (COVID-19) situation in order to protect our nation. DHS is working very closely with the Centers for Disease Control and Prevention (CDC), other federal agencies, and public health officials to implement public health control measures related to travelers and materials crossing our borders from the affected regions.

Based on the response to a similar product generated in 2014 in response to the Ebolavirus outbreak in West Africa, the DHS Science and Technology Directorate (DHS S&T) developed the following “master question list” that quickly summarizes what is known, what additional information is needed, and who may be working to address such fundamental questions as, “What is the infectious dose?” and “How long does the virus persist in the environment?” The Master Question List (MQL) is intended to quickly present the current state of available information to government decision makers in the operational response to COVID-19 and allow structured and scientifically guided discussions across the federal government without burdening them with the need to review scientific reports, and to prevent duplication of efforts by highlighting and coordinating research.

The information contained in the following table has been assembled and evaluated by experts from publicly available sources to include reports and articles found in scientific and technical journals, selected sources on the internet, and various media reports. It is intended to serve as a “quick reference” tool and should not be regarded as comprehensive source of information, nor as necessarily representing the official policies, either expressed or implied, of the DHS or the U.S. Government. DHS does not endorse any products or commercial services mentioned in this document. All sources of the information provided are cited so that individual users of this document may independently evaluate the source of that information and its suitability for any particular use. This document is a “living document” that will be updated as needed when new information becomes available.

Table of Contents

Infectious Dose – How much agent will make a healthy individual ill?	3
<p>The human infectious dose of severe acute respiratory syndrome coronavirus 2 (SARS-CoV-2) is unknown by all exposure routes. SARS-CoV-2 is the cause of coronavirus disease 19 (COVID-19). Understanding the infectious dose in humans, for each of the routes that humans may become infected, is critical to predicting the risk the virus poses in general, and to inform decisions on disinfection and decontamination. Animal studies are a plausible surrogate.</p>	
Transmissibility – How does it spread from one host to another? How easily is it spread?	4
<p>SARS-CoV-2 is passed easily between humans, likely through close contact with relatively large droplets and possibly through smaller aerosolized particles. Individuals can transmit SARS-CoV-2 to others before they have symptoms. Undetected cases play a major role in transmission, and most cases are not reported. Identifying the contribution of asymptomatic or pre-symptomatic transmission is important for implementing control measures. Additionally, the relative contribution of different infection sources – fomites, droplets, aerosols, and potentially feces – is unknown.</p>	
Host Range – How many species does it infect? Can it be transferred from species to species?	6
<p>SARS-CoV-2 is closely related to other coronaviruses circulating in bats in Southeast Asia. Previous coronaviruses have passed through an intermediate mammal host before infecting humans. The identity of the SARS-CoV-2 intermediate host is unknown. SARS-CoV-2 uses the same receptor for cell entry as the SARS-CoV-1 coronavirus that circulated in 2002/2003. To date, ferrets, hamsters, cats, and primates have been shown to be susceptible to SARS-CoV-2 infection. Cats can transmit infection to other cats. It is unknown whether these animals can transmit infection to humans. Several animal models have been developed to recreate human-like illness, though to date they have been infected with high dose exposures. Lower dose studies may better replicate human disease acquisition.</p>	
Incubation Period – How long after infection do symptoms appear? Are people infectious during this time?	7
<p>The majority of individuals develop symptoms within 14 days of exposure. For most people, it takes at least 2 days to develop symptoms, and on average symptoms develop 5 days after exposure. Some individuals never develop symptoms but can still transmit disease. While the incubation period is well-characterized, less is known about how long individuals are infectious before, during, and after symptoms. Additionally, the possibility of reinfection warrants more research.</p>	
Clinical Presentation – What are the signs and symptoms of an infected person?	8
<p>Most COVID-19 cases are mild, but severe disease can be found in any age group. Older individuals and those with underlying medical conditions are at higher risk of serious illness and death. Current modeling suggests the overall case fatality rate (CFR) of COVID-19 is approximately 2.4%,¹⁷ but varies substantially by patient age and underlying comorbidities. Evidence suggests that African Americans are at elevated risk of severe symptoms. Additional data on vulnerable subpopulations is needed. Children of all ages are susceptible to COVID-19,⁸⁵ though generally show milder^{57,155} or no symptoms. The true case fatality rate is unknown, as the exact number of cases is uncertain. Testing priorities and case definitions vary by location.</p>	
Protective Immunity – How long does the immune response provide protection from reinfection?	9
<p>Infected patients show productive immune responses, however, more data is needed. Currently, there is no evidence that recovered patients can be reinfected with SARS-CoV-2. Understanding the duration of protective immunity is limited by small sample sizes. Animal models are plausible surrogates. Additional research to quantify the risk of reinfection after weeks, months, and years is needed.</p>	
Clinical Diagnosis – Are there tools to diagnose infected individuals? When during infection are they effective?	10
<p>Diagnosis relies on identifying the genetic signature of the virus in patient nose, throat, or sputum samples. These tests are relatively accurate. Confirmed cases are still underreported. Validated serological (antibody) assays are being developed to help determine who has been exposed to SARS-CoV-2.</p>	

In general, PCR tests appear to be sensitive and specific, though confirmation of symptoms via chest CT is recommended. The efficacy of serological testing should be confirmed.

Pharmaceutical Interventions – Are there effective treatments? Vaccines?.....11

Treatment for COVID-19 is primarily supportive care including ventilation if necessary.^{104, 163} Over 332⁴ clinical trials are ongoing, but results are preliminary.^{66, 28} Convalescent sera is being tested at multiple sites across the US.¹⁶⁴

Work is ongoing to develop a SARS-CoV-2 vaccine in human and animal trials. No preliminary results are available.

In general, the efficacy of various therapeutic options for COVID-19 is unknown, though clinical trial results are beginning to be released.

Non-pharmaceutical Interventions – Are public health control measures effective at reducing spread?.....12

Broad-scale control measures such as stay-at-home orders are effective at reducing movement, and modeling shows evidence that they reduce transmission.

The effect of relaxing control measures is unknown, and research is needed to help plan for easing of restrictions.

As US states have implemented differing control measures at various times, a comprehensive analysis of social distancing efficacy has not yet been conducted.

Environmental Stability – How long does the agent live in the environment?.....13

SARS-CoV-2 can persist on surfaces for at least 3 days and on the surface of a surgical mask for up to 7 days depending on conditions. If aerosolized intentionally, SARS-CoV-2 is stable for at least several hours. The seasonality of COVID-19 transmission is unknown.

Additional testing on SARS-CoV-2, as opposed to surrogate viruses, is needed to support initial estimates of stability.

Decontamination – What are effective methods to kill the agent in the environment?14

Soap and water, as well as common alcohol and chlorine-based cleaners, hand sanitizers, and disinfectants are effective at inactivating SARS-CoV-2 on hands and surfaces.

Methods for decontaminating N95 masks have been approved by the FDA under an Emergency Use Authorization (EUA).

Additional decontamination studies, particularly with regard to PPE and other items in short supply, are needed.

PPE – What PPE is effective, and who should be using it?15

The effectiveness of PPE for SARS-CoV-2 is currently unknown, and data from other related coronaviruses are used for guidance. Healthcare workers are at high risk of acquiring COVID-19, even with recommended PPE.

Most PPE recommendations have not been made based on SARS-CoV-2 data, and comparative efficacy of different PPE for different tasks (e.g., intubation) is unknown. Identification of efficacious PPE for healthcare workers is critical due to their high rates of infection.

Forensics – Natural vs intentional use? Tests to be used for attribution.16

All current evidence supports the natural emergence of SARS-CoV-2 via a bat and possible intermediate mammal species.

Identifying the intermediate species between bats and humans would aid in reducing potential spillover from a natural source.

Genomics – How does the disease agent compare to previous strains?17

Current evidence suggests that SARS-CoV-2 accumulates substitutions and mutations at a rate similar to other coronaviruses. Mutations and deletions in specific portions of the SARS-CoV-2 genome have not been linked to any changes in transmission or disease severity, though modeling work is attempting to identify possible changes.

Research linking genetic changes to differences in phenotype (e.g., transmissibility, virulence, progression in patients) is needed.

SARS-CoV-2 (COVID-19)	Infectious Dose – How much agent will make a healthy individual ill?
<p>What do we know?</p>	<p>The human infectious dose of severe acute respiratory syndrome coronavirus 2 (SARS-CoV-2) is unknown by all exposure routes. SARS-CoV-2 is the cause of coronavirus disease 19 (COVID-19). <i>Work using SARS-CoV-2</i></p> <ul style="list-style-type: none"> • A total dose of approximately 700,000 plaque-forming units (PFU) of the novel coronavirus SARS-CoV-2 infected cynomolgus macaques via combination intranasal and intratracheal exposure (10^6 TCID₅₀ total dose).²⁰¹ Macaques did not exhibit clinical symptoms, but virus was shed from the nose and throat.²⁰¹ • Rhesus macaques are effectively infected with SARS-CoV-2 via the ocular conjunctival and intratracheal route at a dose of approximately 700,000 PFU (10^6 TCID₅₀).⁸³ • Rhesus macaques infected with 2,600,000 TCID₅₀ of SARS-CoV-2 by the intranasal, intratracheal, oral and ocular routes combined recapitulate moderate disease observed in the majority of human cases.¹⁷⁰ • Ferrets infected with 316,000 TCID₅₀¹²⁴ or 600,000 TCID₅₀¹⁹⁵ of SARS-CoV-2 by the intranasal route show similar symptoms to human disease.^{124, 195} Uninfected ferrets in direct contact with infected ferrets test positive and show disease as early as 2 days post-contact.¹²⁴ <i>In one study, direct contact was required to transfer infection between ferrets,¹²⁴ however, transmission without direct contact was found in another study.¹⁹⁵</i> • Syrian Golden Hamsters infected with 100,000 PFU via the intranasal route closely resemble human respiratory infection. Uninfected hamsters in close contact with infected hamsters show symptoms within 4 days of exposure.⁵³ • Domestic cats exposed to 100,000 PFU of SARS-CoV-2 via the intranasal route developed severe pathological symptoms including lesions in the nose, throat, and lungs.²¹³ Juvenile cats exhibited more severe symptoms than subadults.²¹³ <p><i>Related Coronaviruses</i></p> <ul style="list-style-type: none"> • The infectious dose for severe acute respiratory syndrome (SARS) coronavirus 1 (SARS-CoV-1) in mice is estimated to be between 67-540 PFU (average 240 PFU, intranasal route).^{80, 82} • Genetically modified mice exposed intranasally to doses of Middle East respiratory syndrome (MERS) coronavirus (MERS-CoV) between 100 and 500,000 PFU show signs of infection. Infection with higher doses result in severe syndromes.^{13, 67, 141, 263}
<p>What do we need to know?</p>	<p>Identifying the infectious dose for humans by the various routes through which individuals become infected is critical to the effective development of computational models to predict risk, develop diagnostics and countermeasures, and effective decontamination strategies. Animal studies are a plausible surrogate.</p> <ul style="list-style-type: none"> • Human infectious dose by aerosol route • Human infectious dose by surface contact (fomite) • Human infectious dose by fecal-oral route

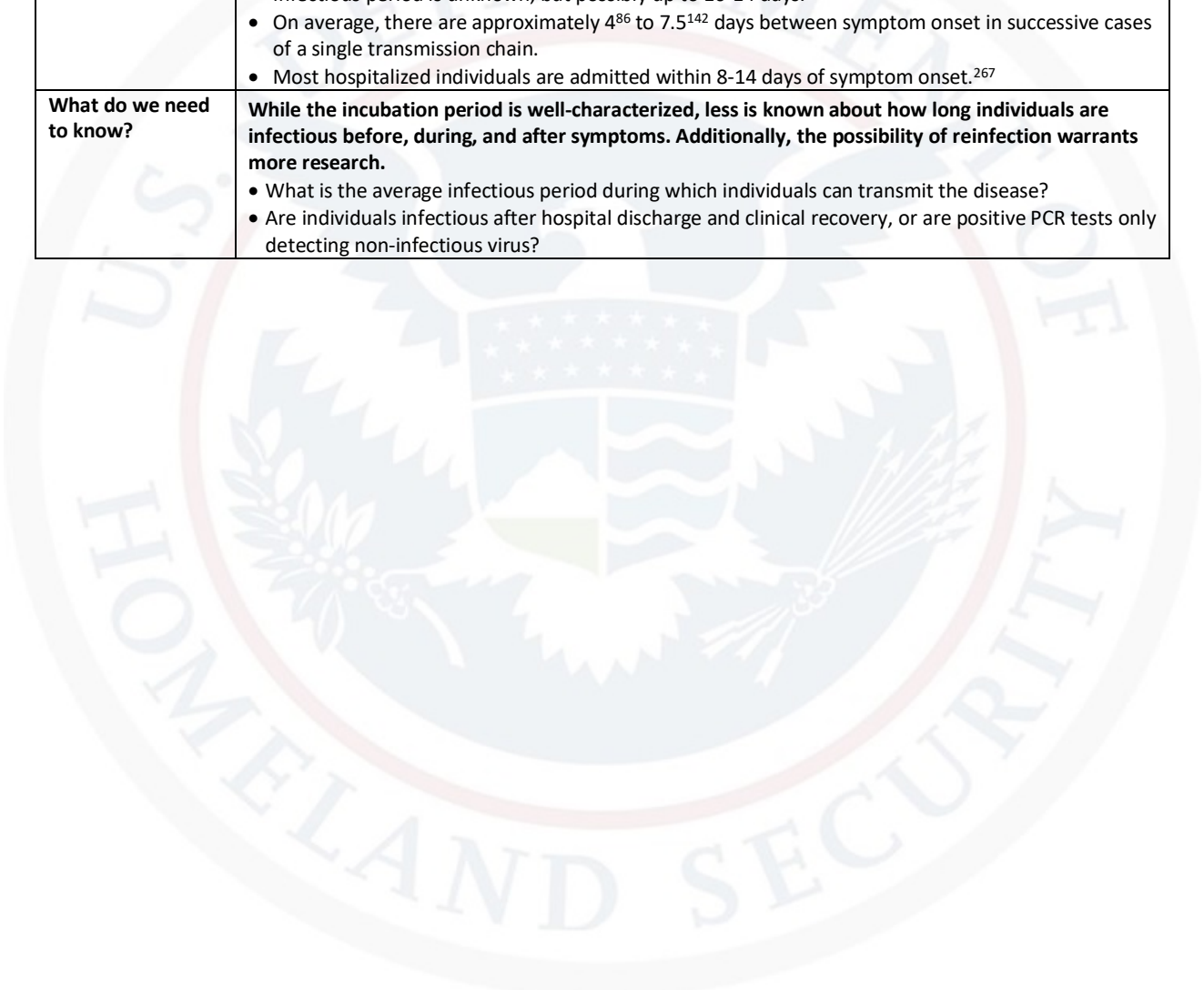
SARS-CoV-2 (COVID-19)	Transmissibility – How does it spread from one host to another? How easily is it spread?
<p>What do we know?</p>	<p>SARS-CoV-2 is passed easily between humans, likely through close contact with relatively large droplets and possibly through smaller aerosolized particles.</p> <ul style="list-style-type: none"> • Pandemic COVID-19 has caused 2,501,156 infections and 171,810 deaths¹¹⁸ in at least 185 countries and territories (as of 4/21/2020).^{44, 209, 241} • In the US there are 788,920 confirmed SARS-CoV-2 cases across all 50 US states, with 42,458 deaths (as of 4/21/2020).¹¹⁸ • High-quality estimates of human transmissibility (R_0) range from 2.2 to 3.1.^{159, 183, 197, 249, 262} • SARS-CoV-2 is believed to spread through close contact and droplet transmission,⁴⁸ with fomite transmission likely¹¹⁹ and close-contact aerosol transmission plausible^{34, 103} but unconfirmed.²⁴⁰ • Aerosolized virus has been detected in COVID-19 patient rooms, with particle sizes within the human respirable range (0.25 – 2.5 μm).¹⁵¹ • Extensive contamination of patient rooms indicates the potential for airborne transmission, though to date infectious virus has not been isolated from aerosol samples.²⁰⁶ Contamination may be worse in intensive care rooms, with viral RNA detected up to 4 meters from patient beds.¹⁰⁶ • Limited evidence suggests that SARS-CoV-2 may be spread by conversation and exhalation in the absence of cough, however more work is needed.^{7, 11, 140, 206} • Experimentally infected ferrets were able to transmit SARS-CoV-2 to other ferrets by both direct contact (another ferret in same enclosure) as well as through the air (ferrets in an adjacent enclosure, separated by 10 cm).¹⁹⁵ <p>Individuals can transmit SARS-CoV-2 to others before they have symptoms.</p> <ul style="list-style-type: none"> • SARS-CoV-2 replicates in the upper respiratory tract (e.g., throat), and infectious virus is detectable in throat and lung tissue for at least 8 days.²⁴⁴ • Pre-symptomatic²⁶⁵ or asymptomatic²² patients can transmit SARS-CoV-2. 12%⁸⁶ of all cases are estimated to be due to asymptomatic transmission, and between 23%¹⁵⁰ and 44%¹⁰⁷ of infections may be caused by pre-symptomatic transmission. • Individuals may be infectious for 1-3 days prior to symptom onset.²³³ • Severe cases are more likely to transmit disease, and most new infections are within households of infected patients.¹⁵⁶ In China, it is estimated that infected individuals transmit COVID-19 to 16.7% of their household contacts.¹⁴⁴ • Asymptomatic individuals are estimated to be infectious for a median of 9.5 days.¹¹³ <p>Undetected cases play a major role in transmission, and most cases are not reported.</p> <ul style="list-style-type: none"> • Models suggest up to 86% of early COVID-19 cases in China were undetected, and these infections were the source for 79% of reported cases.¹⁴³ • Models estimate that the true number of cases may be approximately 11 times greater than the reported number of cases in the UK,²⁵⁸ and 5 to 10 times greater than the reported number of cases in the US.¹²⁰ • Assuming a case fatality rate of 1.4%, researchers estimate that only 12% of cases in the US are confirmed and reported.²⁰⁴ • Preliminary estimates of the case reporting rate vary widely among countries, from roughly 1 reported case for every 3 actual cases (in Germany), to 1 in 149 (in China).¹²⁸ Additional estimates of the level of reporting are needed. • In Ohio, every inmate at each of three prisons was tested for COVID-19 regardless of symptoms. Rates of infection were 21%, 58%, and 78%. The percentage of inmates that presented with symptoms is unknown.⁷³

SARS-CoV-2 (COVID-19)	Transmissibility – How does it spread from one host to another? How easily is it spread?
<p>What do we need to know?</p>	<p>Identifying the contribution of asymptomatic or pre-symptomatic transmission is important for implementing control measures. Additionally, the relative contribution of different infection sources – fomites, droplets, aerosols, and potentially feces – is unknown.</p> <ul style="list-style-type: none"> • Capability of SARS-CoV-2 to be transmitted by contact with fomites (phones, doorknobs, surfaces, clothing, etc.) – see also Experimental Stability • Updated person to person transmission rates (e.g., R_0) as control measures take effect. • Can individuals become re-infected with SARS-CoV-2? • Is the R_0 estimate higher in healthcare or long-term care facilities? • When will infections peak in various cities and countries? • Are small aerosol exposures capable of infecting humans? • How far do infectious small aerosols travel?



SARS-CoV-2 (COVID-19)	Host Range – How many species does it infect? Can it transfer from species to species?
<p>What do we know?</p>	<p>SARS-CoV-2 is closely related to other coronaviruses circulating in bats in Southeast Asia. Previous coronaviruses have passed through an intermediate mammal host before infecting humans, but the identity of the SARS-CoV-2 intermediate host is unknown.</p> <ul style="list-style-type: none"> • Early genomic analysis indicates similarities to SARS-CoV-1,²⁶⁸ with a suggested bat origin.^{5,68, 268} • Positive samples from the South China Seafood Market strongly suggests a wildlife source,⁵⁰ though it is possible that the virus was circulating in humans before the disease was associated with the seafood market.^{27, 70, 252, 260} • Analysis of SARS-CoV-2 genomes suggests that a non-bat intermediate species is responsible for the beginning of the outbreak.²⁰⁰ The identity of the intermediate host remains unknown.^{145, 147-148} • Viruses similar to SARS-CoV-2 were present in pangolin samples collected several years ago.¹³² <p>SARS-CoV-2 uses the same receptor for cell entry as the SARS-CoV-1 coronavirus that circulated in 2002/2003.</p> <ul style="list-style-type: none"> • Experiments show that SARS-CoV-2 Spike (S) receptor-binding domain binds to the human cell receptor (ACE2) stronger than SARS-CoV-1,²⁴⁷ potentially explaining its high transmissibility. The same work suggests that differences between SARS-CoV-2 and SARS-CoV-1 Spike proteins may limit the therapeutic ability of SARS antibody treatments.²⁴⁷ • Modeling of SARS-CoV-2 Spike and ACE2 proteins suggests that SARS-CoV-2 can bind and infect human, bat, civet, monkey and swine cells.²²⁷ • Genetic and protein analysis of primates suggests that the apes and monkeys from Africa and Asia share the same amino acids as the human ACE2 receptor, indicating plausible susceptibility to SARS-CoV-2 infection.¹⁶⁵ This reiterates the importance of identifying potential SARS-CoV-2 animal reservoirs. • Receptor binding is not the only feature of coronaviruses that facilitate cell entry, however, changes in proteolytic cleavage of Spike protein can also affect animal host range.¹⁶⁶ <p>To date, ferrets, hamsters, cats, and primates have been shown to be susceptible to SARS-CoV-2 infection. Cats can transmit infection to other cats. It is unknown whether these animals can transmit infection to humans.</p> <ul style="list-style-type: none"> • Animal model studies suggest that Golden Syrian hamsters, primates, and ferrets may be susceptible to infection.^{53, 124} • Domestic cats are susceptible to infection with SARS-CoV-2 (100,000 PFU via the intranasal route), and can transmit the virus to other cats via droplet or short-distance aerosol.²¹³ Dogs exposed to SARS-CoV-2 showed limited evidence of infection, producing anti-SARS-CoV-2 antibodies but exhibited no clinical symptoms.²¹³ • Wild cats (tigers)²³² can be infected with SARS-CoV-2, although their ability to spread to humans is unknown.^{161, 261} • Ducks, chickens, and pigs remained uninfected after experimental SARS-CoV-2 inoculation (30,000 CFU for ducks and chickens, 100,000 PFU for pigs, all via intranasal route).²¹³ There is currently no evidence that SARS-CoV-2 infects livestock.¹¹⁶
<p>What do we need to know?</p>	<p>Several animal models have been developed to recreate human-like illness, though to date they have been infected with high dose exposures. Lower dose studies may better replicate human disease acquisition.</p> <ul style="list-style-type: none"> • What is the intermediate host(s)? • What are the mutations in SARS-CoV-2 that allowed human infection and transmission? • What other animals can SARS-CoV-2 infect (e.g., pet cats and dogs, potential wildlife reservoirs)? • Can infected animals transmit to humans (e.g., pet cats and dogs to humans)?

SARS-CoV-2 (COVID-19)	Incubation period – How long after infection do symptoms appear? Are people infectious during this time?
<p>What do we know?</p>	<p>The majority of individuals develop symptoms within 14 days of exposure. For most people, it takes at least 2 days to develop symptoms, and on average symptoms develop 5 days after exposure. Some individuals never develop symptoms but can still transmit disease.</p> <ul style="list-style-type: none"> • The best current estimate of the COVID-19 incubation period is 5.1 days, with 99% of individuals exhibiting symptoms within 14 days of exposure.¹³⁶ Fewer than 2.5% of infected individuals show symptoms sooner than 2 days after exposure.¹³⁶ • Individuals can test positive for COVID-19 even if they lack clinical symptoms.^{22, 52, 104, 218, 265} • Individuals can be infectious while asymptomatic,^{48, 202, 218, 265} and asymptomatic individuals can have similar amounts of virus in their nose and throat as symptomatic individuals.²⁷⁰ • Peak infectiousness may be during the incubation period one day before symptoms develop.¹⁰⁷ • Infectious period is unknown, but possibly up to 10-14 days.^{10, 143, 209} • On average, there are approximately 4⁸⁶ to 7.5¹⁴² days between symptom onset in successive cases of a single transmission chain. • Most hospitalized individuals are admitted within 8-14 days of symptom onset.²⁶⁷
<p>What do we need to know?</p>	<p>While the incubation period is well-characterized, less is known about how long individuals are infectious before, during, and after symptoms. Additionally, the possibility of reinfection warrants more research.</p> <ul style="list-style-type: none"> • What is the average infectious period during which individuals can transmit the disease? • Are individuals infectious after hospital discharge and clinical recovery, or are positive PCR tests only detecting non-infectious virus?



SARS-CoV-2 (COVID-19)	Clinical presentation – What are the signs and symptoms of an infected person?
<p>What do we know?</p>	<p>Most COVID-19 cases are mild, but severe disease can be found in any age group. Older individuals and those with underlying medical conditions are at higher risk of serious illness and death.</p> <ul style="list-style-type: none"> • The majority of COVID-19 cases are mild (81%, n=44,000 cases).²¹⁸ • Initial COVID-19 symptoms include fever (87.9% overall, but only 44-52% present with fever initially),^{18, 104} cough (67.7%),¹⁰⁴ fatigue, shortness of breath, headache, and reduced lymphocyte count.^{49, 56, 114} Headache is uncommon.⁵⁵ Diarrhea may be uncommon,^{114, 142} though lack of appetite may be an early symptom.¹⁸¹ • Complications include acute respiratory distress (ARDS, 17-29% of hospitalized patients, leading to death in 4-15% of cases),^{60, 114, 228} pneumonia,¹⁸⁰ cardiac injury (20%),²¹⁴ secondary infection, kidney failure, arrhythmia, sepsis, and shock.^{104, 114, 228, 267} • Most deaths are caused by respiratory failure or respiratory failure combined with myocardial (heart) damage.²⁰³ • Approximately 15% of hospitalized patients are classified as severe,^{104, 218} and approximately 5% of patients are admitted to the ICU.^{104, 218} Of the patients requiring mechanical ventilation, 70% required supplemental oxygen upon hospital admission, suggesting rapid deterioration with respiratory failure.¹⁰¹ • Loss of taste and smell appears in 5-30% of patients who test positive, however, approximately 18% of individuals who test negative also report this symptom. Survey results indicate higher self-reported rates of smell and taste loss in COVID-19 positive patients (up to 68% and 71%, respectively), and only 16%-17% self-reported in COVID-19 negative patients.²⁵⁶ More work is needed.^{21, 65, 98} • Several studies suggest that SARS-CoV-2 is not transmitted from mother to child during birth,^{59, 61, 208, 257} however larger studies are needed. • Ocular symptoms such as conjunctivitis have been seen in severe COVID-19 cases,²⁵¹ and blood clots appear more common in COVID-19 patients in intensive care than the general population.¹²⁶ • Kidney damage may be relatively common in patients with severe COVID-19, and there is evidence of SARS-CoV-2 presence in kidney tissue.²¹⁵ • Up to 67% of patients with clinically asymptomatic infection may still show CT evidence of pneumonia.²³⁰ • Almost 50% of hospitalized patients in the US have either hypertension or obesity as comorbidities, and approximately 25% experienced diarrhea or nausea and vomiting.⁹⁷ <p>Current modeling suggests the overall case fatality rate (CFR) of COVID-19 is approximately 2.4%,¹⁷ but varies substantially by patient age and underlying comorbidities.</p> <ul style="list-style-type: none"> • The CFR depends on comorbidities, and cardiovascular disease, hypertension, diabetes, and respiratory conditions all increase the CFR.^{218, 267} • The CFR increases with age, and individuals >60 are at higher risk of death.^{218, 267} In the US, 34% of hospitalizations have been individuals younger than 44 years old.⁸ Over >60% of confirmed fatalities have been male.²¹⁸ • Variation in the CFR between countries may be due to demographics, testing criteria, and how COVID-19 related deaths are defined.¹⁷⁵ <p>Evidence suggests that African Americans are at elevated risk of severe symptoms. Additional data on vulnerable subpopulations is needed.</p> <ul style="list-style-type: none"> • A review of US COVID-19 patients revealed that African Americans are disproportionately represented in hospitalized populations (comprising 33% of hospitalized patients compared to only 18% of the base study population).⁹⁷ Additional research highlighting potentially vulnerable subpopulations is needed. <p>Children of all ages are susceptible to COVID-19,⁸⁵ though generally show milder^{57, 155} or no symptoms.</p> <ul style="list-style-type: none"> • Up to 28% of children may be asymptomatic.¹⁸⁶ • Severe symptoms in children are possible,¹⁴⁹ and infant deaths have been recorded.^{36, 155}
<p>What do we need to know?</p>	<p>The true case fatality rate is unknown, as the exact number of cases is uncertain. Testing priorities and case definitions vary by location.</p> <ul style="list-style-type: none"> • How long does it take for infected individuals to recover outside of a healthcare setting? • Are pregnant women at greater risk of complications during labor?¹⁴⁶

SARS-CoV-2 (COVID-19)	Protective Immunity – How long does the immune response provide protection from reinfection?
<p>What do we know?</p>	<p>Infected patients show productive immune responses, however more data is needed.</p> <ul style="list-style-type: none"> • In a limited study (n=9), hospitalized patients shed high levels of infectious virus for 7 days via the nasal-pharyngeal route, 50% of patients produced antibodies within 7 days, and all patients produced antibodies by 14 days. Antibody production did not correlate with lower viral load.²⁴⁴ • In a larger study (n=175), most patients developed neutralizing antibodies within 10-15 days after disease onset. Elderly patients had significantly higher neutralizing antibody titers than younger patients.²⁴⁸ • Based on one patient, a productive immune response was generated and sustained for at least 7 days.²¹⁹ Previous studies on coronavirus immunity suggest that neutralizing antibodies may wane after several years.^{38, 250} More data is needed. • A small subset of COVID-19 patients in China (8%) did not develop a serological response to infection, however, the potential for reinfection in these patients is unknown.²⁴⁸ Interestingly, the majority of patients that failed to develop a quantifiable immune response was < 40 years old. <p>Currently, there is no evidence that recovered patients can be reinfected with SARS-CoV-2.</p> <ul style="list-style-type: none"> • Experimentally infected macaques were not capable of being reinfected after their primary infection resolved.²⁴ • According to the WHO, there is no evidence of re-infection with SARS-CoV-2 after recovery.¹³⁵ • Patients can test positive via PCR for up to 37 days after symptoms appear,²⁶⁷ and after recovery and hospital discharge.¹³³ The ability of these individuals to infect others is unknown.
<p>What do we need to know?</p>	<p>Understanding the duration of protective immunity is limited by small sample sizes. Animal models are plausible surrogates. Additional research to quantify the risk of reinfection after weeks, months, and years is needed.</p> <ul style="list-style-type: none"> • How long does the immune response last? • Is there evidence of waning immunity? • Can humans become reinfected? • Are patients who test positive weeks after discharge from hospital capable of transmitting infection? • How does the patient immune response vary by age or disease severity?

SARS-CoV-2 (COVID-19)	Clinical Diagnosis – Are there tools to diagnose infected individuals? When during infection are they effective?
<p>What do we know?</p>	<p>Diagnosis relies on identifying the genetic signature of the virus in patient nose, throat, or sputum samples. These tests are relatively accurate. Confirmed cases are still underreported.</p> <ul style="list-style-type: none"> • US CDC has expanded patient testing criteria to include symptomatic patients at clinician discretion.²⁶ • PCR protocols and primers have been widely shared internationally.^{43, 72, 142, 212, 237, 242} PCR-based diagnostic assays are unable to differentiate between active and inactive virus. • Broad testing in Iceland suggests that approximately 50% of those who test positive are symptom-free at the time of testing.^{19, 168} • A combination of pharyngeal (throat) RT-PCR and chest tomography are the most effective diagnostic criteria (correctly diagnose 91.9% of infections).¹⁹³ A single throat swab detects 78.2% of infections, and duplicate tests identify 86.2% of infections.¹⁹³ • Nasal and pharyngeal swabs may be less effective as diagnostic specimens than sputum and bronchoalveolar lavage fluid,²²⁹ although recent evidence suggests this may not always be the case.²⁴⁴ More work is needed. • RT-PCR tests can identify asymptomatic cases, and SARS-CoV-2 infection was identified in 2/114 individuals cleared by clinical assessment.¹¹² • Combination RT-PCR and serology (antibody) testing may increase the ability to diagnose patients with mild symptoms, or identify patients at higher risk of severe disease.²⁶⁴ • The FDA released an Emergency Use Authorization enabling laboratories to develop and use tests in-house for patient diagnosis.⁹⁴ • Updated tests from the US CDC are available to states.^{43, 48} • Multiple rapid or real-time test kits have been produced by universities and industry, including the Wuhan Institute of Virology,⁷⁶ BGI,³⁰ Cepheid,²²⁶ Abbot,⁹² and Mesa Biotech.³¹ • The US CDC is developing serological tests to determine what proportion of the population has been exposed to SARS-CoV-2.¹²² A rapid antibody test by Cellex is now authorized by the FDA.^{111, 235} • Home tests are being developed, however none are FDA approved, nor are they useable as a diagnostic.^{171-172, 182} • Interleukin-6 levels of ≥ 80 pg/mL were associated with respiratory failure in a small study (n=41).¹⁰⁸ More work is needed. <p>Validated serological (antibody) assays are being developed to help determine who has been exposed to SARS-CoV-2.</p> <ul style="list-style-type: none"> • Researchers have tested a variety of enzyme-linked immunosorbent assays (ELISA) to determine their sensitivity and specificity to SARS-CoV-2 as well as other coronaviruses. Results show high specificity, though sample sizes for SARS-CoV-2 patients were small.¹⁷⁴ • In one German town, serological testing has been used to identify the percent of the population already exposed to SARS-CoV-2 (14%), which can assist in public health response planning.¹⁹¹ • A preliminary study in Santa Clara, California used serological testing to estimate that between 2.5-4.1% of the population has already been exposed to SARS-CoV-2 since the first confirmed cases in January.²⁹ The study population (n=3,330), however, was not a random sample, potentially biasing estimates upwards.²⁹ Additionally, the false positive rate of the diagnostic assay used may account for a substantial portion of the reported infections.²⁹ • Preliminary, unpublished serological results from Los Angeles, California, also suggest an underlying infection rate of 4.1% (n=896), approximately 55 times larger than the number of reported cases;¹⁶² false positives are still a potential issue. Results from larger, randomized serological surveys are necessary to infer population-level trends. Additionally, positive serological tests do not necessarily indicate the presence of neutralizing antibodies.
<p>What do we need to know?</p>	<p>In general, PCR tests appear to be sensitive and specific, though confirmation of symptoms via chest CT is recommended. The efficacy of serological testing should be confirmed.</p> <ul style="list-style-type: none"> • Eclipse phase of infection (time between infection and detectable disease) in an individual • With limited testing in many locations, how accurate are clinical diagnoses compared to genetic tests? • How effective are different swab specimens as diagnostic samples? • How many serological tests need to be done to obtain an accurate picture of underlying exposure?

SARS-CoV-2 (COVID-19)	Pharmaceutical Interventions – Are there effective treatments? Vaccines?
<p>What do we know?</p>	<p>Treatment for COVID-19 is primarily supportive care including ventilation if necessary.^{104, 163} Over 332⁴ clinical trials are ongoing, but results are preliminary.^{28, 66} Convalescent sera is being tested at multiple sites across the US.¹⁶⁴</p> <ul style="list-style-type: none"> • The WHO is tracking >50 potential vaccines,⁶⁹ and has begun two global clinical trials: Solidarity and Discovery¹³⁰ that include remdesivir, hydroxychloroquine and chloroquine, ritonavir/lopinavir, and ritonavir/lopinavir and interferon-beta.¹³⁰ • Remdesivir given to macaques within 12 hours of SARS-CoV-2 inoculation reduced clinical symptoms and viral replication in the lower respiratory tract compared to controls. Viral replication in the upper respiratory tract was not reduced, suggesting a clinical benefit but limited ability to reduce transmission.²⁴³ • Compassionate use of remdesivir in critically ill patients resulted in increases in clinical outcome, based on oxygen-support class, though the data were not compared to a control group.¹⁰² • Anecdotal reports suggest some benefit of remdesivir in humans with severe disease, though the corresponding clinical trial lacks a control group and has not reported final results.⁹⁶ • Limited, preliminary evidence from clinical trials supports the efficacy of favipiravir,⁵⁸ tocilizumab,²⁵³ intravenous immunoglobulin,⁴⁰ and hydroxychloroquine with azithromycin.^{99, 152} <i>Additional work including sufficiently powered clinical trials are necessary to confirm therapeutic efficacy of any of these compounds.</i> • Limited, preliminary evidence shows mixed efficacy of chloroquine alone,³ and no efficacy from combination ritonavir and lopinavir.³⁹ <i>Additional work is necessary to confirm these results.</i> • A study of 181 COVID-19 patients in France found that hydroxychloroquine (600 mg/day) did not reduce the need for intensive care or reduce mortality compared to a control group.¹⁵⁸ Additionally, 9% of the treatment group suffered cardiac arrhythmias that necessitated premature withdrawal of treatment, though it is possible these patients received azithromycin or another antibiotic concurrently.¹⁵⁸ • Favipiravir has been approved to treat COVID-19 in China.¹ • Teams across the USA are testing passive antibody therapy (convalescent serum)⁴¹ to patients (FDA Investigational New Drug approval).⁹³ In a small trial (5 patients),²¹¹ convalescent sera administration was followed by clinical improvement.²¹¹ • Corticosteroids are commonly given to COVID-19 patients²⁶⁷ at risk of ARDS,²⁵⁴ but their use is not recommended by the US CDC.⁴⁵ • Laboratory testing identified 17 repurposed drugs and remdesivir-like nucleoside inhibitors²¹⁰ with significant antiviral activity, however more research is needed to confirm efficacy.²³⁴ • Additional clinical trials involving anti-inflammatory drugs are recruiting patients (interleukin-6 inhibitors, sarilumab² and tocilizumab⁹). • A blood-cleaning device has been approved by the FDA under an Emergency Use Authorization to filter cytokines from severely ill COVID-19 patients.¹⁶⁷ • For patients with ARDS, prone positioning (placing patients on their stomachs and sides rather than their backs) may aid oxygenation and reduce mortality.^{23, 105} <p>Work is ongoing to develop a SARS-CoV-2 vaccine in human and animal trials. No preliminary results are available.</p> <ul style="list-style-type: none"> • Multiple entities are working to produce a SARS-CoV-2 vaccine,¹⁴ including HHS/NIH/NIAID,^{109, 139} CEPI, Moderna Therapeutics, Pfizer,⁸⁹ Gilead Sciences,^{5-6, 173} Sanofi,³² and Johnson and Johnson.¹²¹ Moderna has begun phase 1 clinical vaccine trials.¹⁹⁹
<p>What do we need to know?</p>	<p>In general, the efficacy of various therapeutic options for COVID-19 is unknown, though clinical trial results are beginning to be released.</p> <ul style="list-style-type: none"> • Is the GLS-5000 MERS vaccine²⁵⁹ cross-reactive against SARS-CoV-2? • Efficacy of antibody treatments developed for SARS^{74, 217} and MERS⁵¹ • <i>Are convalescent plasma treatments effective in humans or animals?</i> • What is the efficacy of various MERS and SARS Phase I/II vaccines and other therapeutics? • Are viral replicase inhibitors such as beta-D-N4-hydroxycytidine effective against SARS-CoV-2?²⁵

SARS-CoV-2 (COVID-19)	Non-pharmaceutical Interventions – Are public health control measures effective at reducing spread?
<p>What do we know?</p>	<p>Broad-scale control measures such as stay-at-home orders are effective at reducing movement, and modeling shows evidence that they reduce transmission.</p> <ul style="list-style-type: none"> • Social distancing and other policies are estimated to have reduced COVID-19 spread by 44% in Hong Kong⁷⁵ and reduced spread in China.^{127, 153} • Modeling demonstrates that multifaceted restrictions and quarantines in China reduced the R_0 of SARS-CoV-2 from greater than 3 to less than 1 between January 23rd and February 5th.¹⁷⁹ • Models indicate that a combination of school closures, work restrictions, and other measures are required to effectively limit transmission.⁹⁵ • Preliminary modeling results from Japan suggest that school closures alone were not sufficient to limit COVID-19 spread, though the school closures in questions only applied to students between 6 and 18 years of age.¹¹⁷ • Globally, there is some evidence that implementing social distancing measures has reduced the amount individuals travel, though the data are based on planned rather than actual trips.¹⁶⁰ Mobility in major US cities declined after each public health intervention implemented.¹³⁴ • Restrictive lockdowns in China are estimated to have reduced disease transmission within only a few days.²⁶⁹ • Non-pharmaceutical interventions in China did not reduce transmission equally across all groups; transmission rates in younger individuals, particularly infants, as well as hospital workers continued to increase even while overall transmission rates declined.¹⁷⁹ <p>The effect of relaxing control measures is unknown, and research is needed to help plan for easing of restrictions.</p> <ul style="list-style-type: none"> • Modeling indicates that COVID-19 is likely to become endemic in the US population, with regular, periodic outbreaks, and that additional social or physical distancing measures may be required for several years to keep cases below critical care capacity in absence of a vaccine or effective therapeutic.¹²⁵ Results depend critically on the duration of immunity after exposure.¹²⁵ • Modeling suggests that premature lifting of social distancing measures will substantially increase the number of local COVID-19 cases in Wuhan, China.¹⁸⁴ • Similarly, forecasts in the US estimate a resumption of exponential case growth if social distancing measures are relaxed.⁷⁷ • In the UK, modelers are assessing the efficacy of rolling interventions, whereby social distancing measures are put into place every few weeks to keep healthcare demand below a critical point.²⁵⁸ • A modeling study using Chinese data estimated the impact of relaxing social distancing measures after an initial reduction in disease transmission. Results suggest that if R_0 is allowed to rise above 1, tightening controls may not be enough to keep transmission low; rather, additional effort would be needed to drive R_0 below 1 again, suggesting that carefully balancing control measures to maintain R_0 below 1 would be more efficient than allowing R_0 to increase again in the first place.¹³⁷ • Robust contact tracing and case finding may be needed to control COVID-19 in the US, but would require additional staff and resources to conduct effectively.²³¹ • The WHO released guidelines on a public health strategy, focusing on controlling transmission, ensuring public health capacity is robust, and engaging local communities.²³⁶ • Johns Hopkins released a report outlining how to re-open certain categories of activities (e.g., schools, restaurants, events) while reducing COVID-19 risk. The report also ranks certain activities by their contact intensity, number of contacts, and the potential to modify them to reduce risk.¹⁹⁸
<p>What do we need to know?</p>	<p>As different US states have implemented differing control measures at various times, a comprehensive analysis of social distancing efficacy has not yet been conducted.</p> <ul style="list-style-type: none"> • How many cases in the US have been averted due to social distancing restrictions? • How long does it take for various non-pharmaceutical interventions to show effects? • What are effective surrogate measures of social distancing efficacy (e.g., reduction in travel, contact, traffic, etc.)? • What are plausible options for relaxing social distancing and other intervention measures without resulting in a resurgence of COVID-19 cases?

SARS-CoV-2 (COVID-19)	Environmental Stability – How long does the agent live in the environment?
<p>What do we know?</p>	<p>SARS-CoV-2 can persist on surfaces for at least 3 days and on the surface of a surgical mask for up to 7 days depending on conditions. If aerosolized intentionally, SARS-CoV-2 is stable for at least several hours. The seasonality of COVID-19 transmission is unknown.</p> <p><i>SARS-CoV-2 Data</i></p> <ul style="list-style-type: none"> • SARS-CoV-2 can persist on plastic and metal surfaces between 3 days (21-23°C, 40% RH)²²⁴ and 7 days (22°C, 65% RH). Infectious virus can be recovered from a surgical mask after 7 days (22°C, 65% RH).⁶³ • SARS-CoV-2 has an aerosol half-life of 2.7 hours (particles <5 µm, tested at 21-23°C and 65% RH).²²⁴ • SARS-CoV-2 is susceptible to heat treatment (70°C) but can persist for at least two weeks at refrigerated temperatures (4°C).^{63, 190} • SARS-CoV-2 genetic material (RNA) was detected in symptomatic and asymptomatic cruise ship passenger rooms up to 17 days after cabins were vacated. The infectiousness of this material is not known.¹⁶⁹ <p><i>Surrogate Coronavirus data:</i></p> <ul style="list-style-type: none"> • Studies suggest that other coronaviruses can survive on non-porous surfaces up to 9-10 days (MHV, SARS-CoV),^{42, 54} and porous surfaces for up to 3-5 days (SARS-CoV)⁸⁸ in air conditioned environments (20-25°C, 40-50% RH). • Coronavirus survival tends to be higher at lower temperatures and lower relative humidity (RH),^{42, 54, 188, 225} though infectious virus can persist on surfaces for several days in typical office or hospital conditions.²²⁵ • SARS can persist with trace infectivity for up to 28 days at refrigerated temperatures (4°C) on surfaces.⁴² • No strong evidence exists showing reduction in transmission with seasonal increase in temperature and humidity.¹⁵⁷ • One hour after aerosolization approximately 63% of airborne MERS virus remained viable in a simulated office environment (25°C, 75% RH).¹⁸⁵ • Porous hospital materials, including paper and cotton cloth, maintain infectious SARS-CoV for a shorter time than non-porous material.¹³¹
<p>What do we need to know?</p>	<p>Additional testing on SARS-CoV-2, as opposed to surrogate viruses, is needed to support initial estimates of stability.</p> <ul style="list-style-type: none"> • Stability of SARS-CoV-2 in aerosol, droplets, and other matrices (mucus/sputum, feces) • Particle size distribution (e.g., droplet, large droplet, and true aerosol distribution) • Duration of SARS-CoV-2 infectivity via fomites and surface (contact hazard) • Stability of SARS-CoV-2 on PPE (e.g., Tyvek, nitrile, etc.)

SARS-CoV-2 (COVID-19)	Decontamination – What are effective methods to kill the agent in the environment?
<p>What do we know?</p>	<p>Soap and water, as well as common alcohol and chlorine-based cleaners, hand sanitizers, and disinfectants are effective at inactivating SARS-CoV-2 on hands and surfaces.</p> <p><i>SARS-CoV-2</i></p> <ul style="list-style-type: none"> Alcohol-based hand rubs are effective at inactivating SARS-CoV-2.¹²⁹ Chlorine bleach (1%, 2%), 70% ethanol and 0.05% chlorhexidine are effective against live virus in lab tests.⁶² Twice-daily cleaning with sodium dichloroisocyanurate decontaminated surfaces in COVID-19 patient hospital rooms.¹⁷⁶ EPA has released a list of SARS-CoV-2 disinfectants, but solutions were not tested on live virus.¹² <p><i>Other Coronaviruses</i></p> <ul style="list-style-type: none"> Chlorine-based²³⁹ and ethanol-based⁷¹ solutions are recommended. Heat treatment (56°C) is sufficient to kill coronaviruses,^{188, 266} though effectiveness depends partly on protein in the sample.¹⁸⁸ 70% ethanol, 50% isopropanol, sodium hypochlorite (0.02% bleach), and UV radiation can inactivate several coronaviruses (MHV and CCV).²⁰⁵ Ethanol-based biocides effectively disinfect coronaviruses dried on surfaces, including ethanol containing gels similar to hand sanitizer.^{115, 245} Surface spray disinfectants such as Mikrobac, Dismozon, and Korsolex are effective at reducing infectivity of the closely related SARS-CoV after 30 minutes of contact.¹⁸⁷ Coronaviruses may be resistant to thermal inactivation for up to 7 days when stabilized in stool.²²⁰⁻²²¹ Coronaviruses are more stable in matrixes such as respiratory sputum.⁸⁷ <p>Methods for decontaminating N95 masks have been approved by the FDA under an Emergency Use Authorization (EUA).</p> <ul style="list-style-type: none"> Hydrogen peroxide vapor can repeatedly decontaminate N95 respirators.¹⁹⁶ Devices capable of decontaminating 80,000 masks per day have been granted Emergency Use Authorization from the FDA.⁹⁰ The FDA has issued an Emergency Use Authorization for a system capable of decontaminating 10 N95 masks at a time using devices already present in many US hospitals.³³
<p>What do we need to know?</p>	<p>Additional decontamination studies, particularly with regard to PPE and other items in short supply, are needed.</p> <ul style="list-style-type: none"> What is the minimal contact time for disinfectants? Does contamination with human fluids/waste alter disinfectant efficacy profiles? How effective is air filtration at reducing transmission in healthcare, airplanes, and public spaces? Are landfills and wastewater treatment plants effective at inactivating SARS-CoV-2? Is heat or UV decontamination effective to clean N95 respirators and other types of PPE for multi-use?

SARS-CoV-2 (COVID-19)	PPE – What PPE is effective, and who should be using it?
<p>What do we know?</p>	<p>The effectiveness of PPE for SARS-CoV-2 is currently unknown, and data from other related coronaviruses are used for guidance. Healthcare workers are at high risk of acquiring COVID-19, even with recommended PPE.</p> <ul style="list-style-type: none"> • Healthcare worker illnesses (over 1,000)²¹⁸ demonstrates human-to-human transmission despite isolation, PPE, and infection control.²⁰⁷ • Risk of transmission to healthcare workers appears high, with up to 20% of healthcare workers in Lombardy, Italy becoming infected.¹⁹² • Over 50% of US healthcare workers infected with COVID-19 report work in a healthcare setting as their single source of exposure.³⁷ • Healthcare personnel entering the room [of SARS-CoV-2 patients] should use standard precautions, contact precautions, airborne precautions, and use eye protection (e.g., goggles or a face shield).⁴⁶ • WHO indicates healthcare workers should wear clean long-sleeve gowns as well as gloves.²³⁸ • Respirators (NIOSH-certified N95, EUFFP2 or equivalent) are recommended for those dealing with possible aerosols.²³⁹ Additional protection, such as a Powered Air Purifying Respirator (PAPR) with a full hood, should be considered for high-risk procedures (i.e., intubation, ventilation).³⁵ • Particular attention should be paid to the potential for transmission via exhaled air during supportive respiratory procedures.¹⁰³ • There is evidence both for¹⁵¹ and against¹⁷⁶ the detection of SARS-CoV-2 RNA via air sampling in patient rooms and other hospital areas. • Research at Johns Hopkins Center for Health Security has provided initial estimates of PPE needs in the US: 7.8 billion gloves, 668 million gowns, 360 million surgical masks, and 136 million N95 or similar respirators.²²² • KN95 respirators are, under certain conditions, approved for use under FDA Emergency Use Authorization.⁹¹ <p>Masks may be effective at slowing transmission.</p> <ul style="list-style-type: none"> • Surgical face masks, respirators and homemade face masks may prevent transmission of coronaviruses from infectious individuals (with or without symptoms) to other individuals. ^{79, 138, 223} More work is needed. • On 4/3/2020, the US CDC recommended wearing cloth face masks in public where social distancing measures are difficult to maintain.⁴⁷ • The efficacy of homemade PPE, made with T-shirts, bandanas, or similar materials, is less than standard PPE, but may offer some protection if no other options are available.^{64, 78, 194} • A very small study, involving only 4 patients, found no substantial reduction in physical spread of virus from wearing surgical or cloth masks, and also documented contamination on the inner and outer surfaces of masks.²⁰ Additional work should be done to determine the amount that PPE reduces physical spread of SARS-CoV-2, as the sample size of this study (n=4) was very small.
<p>What do we need to know?</p>	<p>Most PPE recommendations have not been made on SARS-CoV-2 data, and comparative efficacy of different PPE for different tasks (e.g., intubation) is unknown. Identification of efficacious PPE for healthcare workers is critical due to their high rates of infection.</p> <ul style="list-style-type: none"> • What is the importance of aerosol transmission? What is the effective distance of spread via droplet or aerosol? • How effective are barriers such as N95 respirators or surgical masks? • What is the appropriate PPE for first responders? Airport screeners? • What are proper procedures for reducing spread and transmission rates in medical facilities? • How effective are homemade masks at reducing transmission?

SARS-CoV-2 (COVID-19)	Forensics – Natural vs intentional use? Tests to be used for attribution.
<p>What do we know?</p>	<p>All current evidence supports the natural emergence of SARS-CoV-2 via a bat and possible intermediate mammal species.</p> <ul style="list-style-type: none"> • Genomic analysis places SARS-CoV-2 into the beta-coronavirus clade, with a close relationship to bat coronaviruses. The SARS-CoV-2 virus is distinct from SARS-CoV-1 and MERS viruses.⁸⁴ • Genomic analysis suggests that SARS-CoV-2 is a natural variant and is unlikely to be human-derived or otherwise created by “recombination” with other circulating strains of coronavirus.^{15, 268} • Genomic data support at least two plausible origins of SARS-CoV-2: (i) natural selection in a non-human animal host prior to zoonotic transfer, and (ii) natural selection in humans following zoonotic transfer.¹⁵ Both scenarios are consistent with the observed genetic changes found in all known SARS-CoV-2 isolates. • Some SARS-CoV-2 genomic evidence indicates a close relationship with pangolin coronaviruses,²⁴⁶ and data suggests that pangolins may be a natural host for beta-coronaviruses.¹⁴⁷⁻¹⁴⁸ Additional research is needed. • Additionally, “[...] SARS-CoV-2 is not derived from any previously used virus backbone,” reducing the likelihood of laboratory origination,¹⁵ and “[...] genomic evidence does not support the idea that SARS-CoV-2 is a laboratory construct, [though] it is currently impossible to prove or disprove the other theories of its origin.”¹⁵ • Work with other coronaviruses has indicated that heparan sulfate dependence can be an indicator of prior cell passage, due to a mutation in the previous furin enzyme recognition motif.⁸¹
<p>What do we need to know?</p>	<p>Identifying the intermediate species between bats and humans would aid in reducing potential spillover from a natural source.</p> <ul style="list-style-type: none"> • What tests for attribution exist for coronavirus emergence? • What is the identity of the intermediate species? • Are there closely related circulating coronaviruses in bats or other animals with the novel PRRA cleavage site found in SARS-CoV-2?

SARS-CoV-2 (COVID-19)	Genomics – How does the disease agent compare to previous strains?
<p>What do we know?</p>	<p>Current evidence suggests that SARS-CoV-2 accumulates substitutions and mutations at a rate similar to other coronaviruses. Mutations and deletions in specific portions of the SARS-CoV-2 genome have not been linked to any changes in transmission or disease severity, though modeling work is attempting to identify possible changes.</p> <ul style="list-style-type: none"> • There have been no documented cases of SARS-CoV-2 prior to December 2019. • Preliminary genomic analyses, however, suggest that the first human cases of SARS-CoV-2 emerged between 10/19/2019 – 12/17/2019.^{16, 27, 189} • The mutation rate of SARS-CoV-2 is estimated to be similar to other RNA viruses (e.g., SARS, Ebola, Zika), and is currently calculated to be 1.04×10^{-3} substitutions per site per year (n=116 genomes).¹¹⁰ • Pangolin coronaviruses are closely related to both SARS-CoV-2 and closely related bat coronaviruses. Phylogenetic analysis suggests that SARS-CoV-2 is of bat origin, but is closely related to pangolin coronavirus.¹⁴⁷⁻¹⁴⁸ • The SARS-CoV-2 Spike protein, which mediates entry into host cells and is the major determinant of host range, and is very similar to the SARS-CoV-1 Spike protein.¹⁵⁴ The rest of the genome is more closely related to two separate bat¹⁵⁴ and pangolin¹⁴⁸ coronaviruses. • Analysis of SARS-CoV-2 sequences from Singapore has identified a large nucleotide (382 bp) deletion in ORF-8.²¹⁶ The effect of this deletion on transmission or virulence is unknown. • A recent report of virus mutations within patients needs more research.¹²³ Additional analysis of data suggests that the results may be due to experimental methods.^{100, 255} • Structural modeling suggests that specific changes in the genetic sequence of the SARS-CoV-2 Spike protein may enhance binding of the virus to human ACE2 receptors.¹⁷⁷ More specifically, changes to two residues (Q493 and N501) are linked with improving the stability of the virus-receptor binding complex.¹⁷⁷ Additionally, structural modeling identified several mutations found in naturally circulating SARS-CoV-2 genomes that may enhance the stability of the receptor binding domain, potentially increasing binding efficacy.¹⁷⁸ Infectivity assays are needed to validate the genotypic changes and possible phenotypic results identified in these studies.
<p>What do we need to know?</p>	<p>Research linking genetic changes to differences in phenotype (e.g., transmissibility, virulence, progression in patients) is needed.</p> <ul style="list-style-type: none"> • Are there similar genomic differences in the progression of coronavirus strains from bat to intermediate species to human? • Are there different strains or clades of circulating virus? If so, do they differ in virulence?

Table 1. Definitions of commonly-used acronyms

Acronym/ Term	Definition	Description
ACE2	Angiotensin-converting enzyme 2	Acts as a receptor for SARS-CoV, allowing entry into human cells
Airborne transmission	Aerosolization of infectious particles	Aerosolized particles can spread for long distances (e.g., between hospital rooms via HVAC systems)
ARDS	Acute respiratory distress syndrome	Leakage of fluid into the lungs which inhibits respiration and leads to death
Attack rate	Proportion of “at-risk” individuals who develop infection	Defined in terms of “at-risk” population such as schools or households, defines the proportion of individuals in those populations who become infected after contact with an infectious individual
CCV	Canine coronavirus	Canine coronavirus
CFR	Case Fatality Rate	Number of deaths divided by confirmed patients
CoV	Coronavirus	Virus typified by crown-like structures when viewed under electron microscope
COVID-19	Coronavirus disease 19	Official name for the disease caused by the SARS-CoV-2 virus.
Droplet transmission	Sneezing, coughing	Transmission via droplets requires relatively close contact (e.g., within 6 feet)
Fomite	Inanimate vector of disease	Surfaces such as hospital beds, doorknobs, healthcare worker gowns, faucets, etc.
HCW	Healthcare worker	Doctors, nurses, technicians dealing with patients or samples
Incubation period	Time between infection and symptom onset	Time between infection and onset of symptoms typically establishes guidelines for isolating patients before transmission is possible
Infectious period	Length of time an individual can transmit infection to others	Reducing the infectious period is a key method of reducing overall transmission; hospitalization, isolation, and quarantine are all effective methods
Intranasal	Agent deposited into external nares of subject	Simulates inhalation exposure by depositing liquid solution of pathogen/virus into the nose of a test animal, where it is then taken up by the respiratory system.
MERS	Middle-East Respiratory Syndrome	Coronavirus with over 2,000 cases in regional outbreak since 2012
MHV	Mouse hepatitis virus	Coronavirus surrogate
Nosocomial	Healthcare- or hospital-associated infections	Characteristic of SARS and MERS outbreaks, lead to refinement of infection control procedures
PCR	Polymerase chain reaction	PCR (or real-time [RT] or quantitative [Q] PCR) is a method of increasing the amount of genetic material in a sample, which is then used for diagnostic testing to confirm the presence of SARS-CoV-2
PFU	Plaque forming unit	Measurement of the number of infectious virus particles as determined by plaque forming assay. A measurement of sample infectivity.
PPE	Personal protective equipment	Gowns, masks, gloves, and any other measures used to prevent spread between individuals
R ₀	Basic reproduction number	A measure of transmissibility. Specifically, the average number of new infections caused by a typical infectious individual in a wholly susceptible population.
SARS	Severe Acute Respiratory Syndrome	Coronavirus with over 8,000 cases in global 2002-2003 outbreak
SARS-CoV-2	Severe acute respiratory syndrome coronavirus 2	Official name for the virus previously known as 2019-nCoV.

Acronym/ Term	Definition	Description
Serial interval	Length of time between symptom onset of successive cases in a transmission chain	The serial interval can be used to estimate R_0 , and is useful for estimating the rate of outbreak spread
Superspreading	One individual responsible for an abnormally large number of secondary infections	Superspreading can be caused by high variance in the distribution of secondary cases caused by a single individual; most individuals infect very few people, while some infect a large number, even with the same average number of secondary infections
TCID ₅₀	50% Tissue Culture Infectious Dose	The number of infectious units which will infect 50% of tissue culture monolayers. A measurement of sample infectivity.
Transgenic	Genetically modified	In this case, animal models modified to be more susceptible to MERS and/or SARS by adding proteins or receptors necessary for infection



Literature Cited:

1. (U) China approves first anti-viral drug against coronavirus Covid-19. *Pharmaceutical Technology* 2020.
2. (U) Evaluation of the Efficacy and Safety of Sarilumab in Hospitalized Patients With COVID-19. <https://clinicaltrials.gov/ct2/show/NCT04315298?term=sarilumab&cond=covid&draw=2&rank=1>.
3. (U) French researcher posts successful Covid-19 drug trial. *Connexion*: 2020. <https://www.connexionfrance.com/French-news/French-researcher-in-Marseille-posts-successful-Covid-19-coronavirus-drug-trial-results>
4. (U) Global coalition to accelerate COVID-19 clinical research in resource-limited settings. *The Lancet*. [https://doi.org/10.1016/S0140-6736\(20\)30798-4](https://doi.org/10.1016/S0140-6736(20)30798-4)
5. (U) A Multicenter, Adaptive, Randomized Blinded Controlled Trial of the Safety and Efficacy of Investigational Therapeutics for the Treatment of COVID-19 in Hospitalized Adults 2020. <https://clinicaltrials.gov/ct2/show/NCT04280705>
6. (U) Phase I, Open-Label, Dose-Ranging Study of the Safety and Immunogenicity of 2019-nCoV Vaccine (mRNA-1273) in Healthy Adults 2020. <https://clinicaltrials.gov/ct2/show/record/NCT04283461?term=mrna-1273&draw=2&rank=1>
7. (U) *Rapid Expert Consultation on the Possibility of Bioaerosol Spread of SARS-CoV-2 for the COVID-19 Pandemic (April 1, 2020)*. The National Academies Press: Washington, DC, 2020. <https://www.nap.edu/catalog/25769/rapid-expert-consultation-on-the-possibility-of-bioaerosol-spread-of-sars-cov-2-for-the-covid-19-pandemic-april-1-2020>
8. (U) Severe Outcomes Among Patients with Coronavirus Disease 2019 (COVID-19) — United States, February 12–March 16, 2020. *MMWR* 2020. https://www.cdc.gov/mmwr/volumes/69/wr/mm6912e2.htm?s_cid=mm6912e2_w#suggestedcitation
9. (U) A Study to Evaluate the Safety and Efficacy of Tocilizumab in Patients With Severe COVID-19 Pneumonia (COVACTA). <https://clinicaltrials.gov/ct2/show/NCT04320615?term=actemra&cond=covid&draw=2>.
10. (U) [Wuhan Pneumonia] The Hospital Authority stated that 2 critically ill patients needed external life support treatment. <https://www.singtao.ca/4037242/2020-01-14/news-%E3%80%90%E6%AD%A6%E6%BC%A2%E8%82%BA%E7%82%8E%E3%80%91%E9%86%AB%E7%AE%A1%E5%B1%80%E6%8C%872%E5%90%8D%E9%87%8D%E7%97%87%E7%97%85%E6%82%A3%E9%9C%80%E9%AB%94%E5%A4%96%E7%94%9F%E5%91%BD%E6%94%AF%E6%8C%81%E6%B2%BB%E7%99%82/?variant=zh-hk>.
11. (U) AAAS, You may be able to spread coronavirus just by breathing, new report finds. *Science* 2 April, 2020.
12. (U) Agency, U. S. E. P., EPA’s Registered Antimicrobial Products for Use Against Novel Coronavirus SARS-CoV-2, the Cause of COVID-19. <https://www.epa.gov/pesticide-registration/list-n-disinfectants-use-against-sars-cov-2>.
13. (U) Agrawal, A. S.; Garron, T.; Tao, X.; Peng, B. H.; Wakamiya, M.; Chan, T. S.; Couch, R. B.; Tseng, C. T., Generation of a transgenic mouse model of Middle East respiratory syndrome coronavirus infection and disease. *J Virol* 2015, 89 (7), 3659-70. <https://www.ncbi.nlm.nih.gov/pmc/articles/PMC4403411/pdf/zjv3659.pdf>
14. (U) Amanat, F.; Krammer, F., SARS-CoV-2 vaccines: status report. *Journal of Immunology* 2020, Early View. <https://marlin-prod.literatumonline.com/pb-assets/journals/research/immunity/SARS-CoV-2%20vaccines%20status%20report.pdf>
15. (U) Andersen, K. G.; Rambaut, A.; Lipkin, W. I.; Holmes, E. C.; Garry, R. F., The proximal origin of SARS-CoV-2. *Nature Medicine* 2020. <https://doi.org/10.1038/s41591-020-0820-9>

16. (U) Anderson, K., Estimates of the clock and TMRCA for 2019-nCoV based on 27 genomes. <http://virological.org/t/clock-and-tmrca-based-on-27-genomes/347> (accessed 01/26/2020).
17. (U) Angelopoulos, A. N.; Pathak, R.; Varma, R.; Jordan, M. I., Identifying and Correcting Bias from Time- and Severity-Dependent Reporting Rates in the Estimation of the COVID-19 Case Fatality Rate. *arXiv* **2020**. <https://arxiv.org/abs/2003.08592>
18. (U) Arentz, M.; Yim, E.; Klaff, L.; Lokhandwala, S.; Riedo, F. X.; Chong, M.; Lee, M., Characteristics and Outcomes of 21 Critically Ill Patients With COVID-19 in Washington State. *JAMA* **2020**. <https://doi.org/10.1001/jama.2020.4326>
19. (U) Assunção, M., Iceland's coronavirus testing suggests 50% of cases have no symptoms. *NY Daily News* 01 April, 2020.
20. (U) Bae, S.; Kim, M.-C.; Kim, J. Y.; Cha, H.-H.; Lim, J. S.; Jung, J.; Kim, M.-J.; Oh, D. K.; Lee, M.-K.; Choi, S.-H.; Sung, M.; Hong, S.-B.; Chung, J.-W.; Kim, S.-H., Effectiveness of Surgical and Cotton Masks in Blocking SARS-CoV-2: A Controlled Comparison in 4 Patients. *Annals of Internal Medicine* **2020**. <https://doi.org/10.7326/M20-1342>
21. (U) Bagheri, S. H. R.; Asghari, A. M.; Farhadi, M.; Shamshiri, A. R.; Kabir, A.; Kamrava, S. K.; Jalessi, M.; Mohebbi, A.; Alizadeh, R.; Honarmand, A. A.; Ghalehbaghi, B.; Salimi, A., Coincidence of COVID-19 epidemic and olfactory dysfunction outbreak. *medRxiv* **2020**, 2020.03.23.20041889. <http://medrxiv.org/content/early/2020/03/27/2020.03.23.20041889.abstract>
22. (U) Bai, Y.; Yao, L.; Wei, T.; Tian, F.; Jin, D.-Y.; Chen, L.; Wang, M., Presumed Asymptomatic Carrier Transmission of COVID-19. *JAMA*.
23. (U) Bamford, P.; Bentley, A.; Dean, J.; Whitemore, D.; Wilson-Baig, N., *ICS Guidance for the Prone Positioning of the Conscious COVID Patient 2020*; Intensive Care Society: 2020. <https://emcrit.org/wp-content/uploads/2020/04/2020-04-12-Guidance-for-conscious-proning.pdf>
24. (U) Bao, L.; Deng, W.; Gao, H.; Xiao, C.; Liu, J.; Xue, J.; Lv, Q.; Liu, J.; Yu, P.; Xu, Y.; Qi, F.; Qu, Y.; Li, F.; Xiang, Z.; Yu, H.; Gong, S.; Liu, M.; Wang, G.; Wang, S.; Song, Z.; Zhao, W.; Han, Y.; Zhao, L.; Liu, X.; Wei, Q.; Qin, C., Reinfection could not occur in SARS-CoV-2 infected rhesus macaques. *bioRxiv* **2020**, 2020.03.13.990226. <https://www.biorxiv.org/content/biorxiv/early/2020/03/14/2020.03.13.990226.full.pdf>
25. (U) Barnard, D. L.; Hubbard, V. D.; Burton, J.; Smee, D. F.; Morrey, J. D.; Otto, M. J.; Sidwell, R. W., Inhibition of severe acute respiratory syndrome-associated coronavirus (SARSCoV) by calpain inhibitors and beta-D-N4-hydroxycytidine. *Antivir Chem Chemother* **2004**, *15* (1), 15-22. <https://journals.sagepub.com/doi/pdf/10.1177/095632020401500102>
26. (U) BBC, Coronavirus: California declares emergency after death. *BBC* 2020.
27. (U) Bedford, T.; Neher, R., Genomic epidemiology of novel coronavirus (nCoV) using data from GISAID. <https://nextstrain.org/ncov>.
28. (U) Belhadi, D.; Peiffer-Smadja, N.; Yazdanpanah, Y.; Mentré, F.; Laouénan, C., A brief review of antiviral drugs evaluated in registered clinical trials for COVID-19. *medRxiv* **2020**, 2020.03.18.20038190. <https://www.medrxiv.org/content/medrxiv/early/2020/03/20/2020.03.18.20038190.full.pdf>
29. (U) Bendavid, E.; Mulaney, B.; Sood, N.; Shah, S.; Ling, E.; Bromley-Dulfano, R.; Lai, C.; Weissberg, Z.; Saavedra, R.; Tedrow, J.; Tversky, D.; Bogan, A.; Kupiec, T.; Eichner, D.; Gupta, R.; Ioannidis, J.; Bhattacharya, J., COVID-19 Antibody Seroprevalence in Santa Clara County, California. *medRxiv* **2020**, 2020.04.14.20062463. <https://www.medrxiv.org/content/medrxiv/early/2020/04/17/2020.04.14.20062463.full.pdf>
30. (U) BGI, BGI Responds to Novel Coronavirus with Real-Time Detection Kits, Deploys Emergency Team to Wuhan. 2020. <https://www.bgi.com/global/company/news/bgi-responds-to-novel-coronavirus-with-real-time-detection-kits-deploys-emergency-team-to-wuhan/>

31. (U) Biotech, M., Mesa Biotech Receives Emergency Use Authorization from FDA for a 30 Minute Point of Care Molecular COVID-19 Test. Mesa Biotech: 2020.
<https://www.mesabiotech.com/news/euacoronavirus>
32. (U) Branswell, H., Sanofi announces it will work with HHS to develop a coronavirus vaccine. Statnews, Ed. 2020. <https://www.statnews.com/2020/02/18/sanofi-announces-it-will-work-with-hhs-to-develop-coronavirus-vaccine/>
33. (U) Brennan, Z., FDA issues 2nd EUA for decontamination system for N95 masks. *Regulatory Focus* 2020.
34. (U) Brosseau, L. M., COMMENTARY: COVID-19 transmission messages should hinge on science. <http://www.cidrap.umn.edu/news-perspective/2020/03/commentary-covid-19-transmission-messages-should-hinge-science>.
35. (U) Brosseau, L. M.; Jones, R., Commentary: Protecting health workers from airborne MERS-CoV - learning from SARS. <http://www.cidrap.umn.edu/news-perspective/2014/05/commentary-protecting-health-workers-airborne-mers-cov-learning-sars>.
36. (U) Bryner, J., First US infant death linked to COVID-19 reported in Illinois. *LiveScience* 2020.
37. (U) Burrer, S. L.; de Perio, M. A.; Hughes, M. M.; Kuhar, D. T.; Luckhaupt, S. E.; McDaniel, C. J.; Porter, R. M.; Silk, B.; Stuckey, M. J.; Walters, M., Characteristics of health care personnel with COVID-19—United States, February 12–April 9, 2020. **2020**.
38. (U) Callow, K.; Parry, H.; Sergeant, M.; Tyrrell, D., The time course of the immune response to experimental coronavirus infection of man. *Epidemiology & Infection* **1990**, *105* (2), 435-446.
39. (U) Cao, B.; Wang, Y.; Wen, D.; Liu, W.; Wang, J.; Fan, G.; Ruan, L.; Song, B.; Cai, Y.; Wei, M.; Li, X.; Xia, J.; Chen, N.; Xiang, J.; Yu, T.; Bai, T.; Xie, X.; Zhang, L.; Li, C.; Yuan, Y.; Chen, H.; Li, H.; Huang, H.; Tu, S.; Gong, F.; Liu, Y.; Wei, Y.; Dong, C.; Zhou, F.; Gu, X.; Xu, J.; Liu, Z.; Zhang, Y.; Li, H.; Shang, L.; Wang, K.; Li, K.; Zhou, X.; Dong, X.; Qu, Z.; Lu, S.; Hu, X.; Ruan, S.; Luo, S.; Wu, J.; Peng, L.; Cheng, F.; Pan, L.; Zou, J.; Jia, C.; Wang, J.; Liu, X.; Wang, S.; Wu, X.; Ge, Q.; He, J.; Zhan, H.; Qiu, F.; Guo, L.; Huang, C.; Jaki, T.; Hayden, F. G.; Horby, P. W.; Zhang, D.; Wang, C., A Trial of Lopinavir–Ritonavir in Adults Hospitalized with Severe Covid-19. *New England Journal of Medicine* **2020**.
<https://www.nejm.org/doi/full/10.1056/NEJMoa2001282>
40. (U) Cao, W.; Liu, X.; Bai, T.; Fan, H.; Hong, K.; Song, H.; Han, Y.; Lin, L.; Ruan, L.; Li, T., High-dose intravenous immunoglobulin as a therapeutic option for deteriorating patients with Coronavirus Disease 2019. *Open Forum Infectious Diseases* **2020**. <https://doi.org/10.1093/ofid/ofaa102>
41. (U) Casadevall, A.; Pirofski, L.-a., The convalescent sera option for containing COVID-19. *The Journal of Clinical Investigation* **2020**, *130* (4). <https://doi.org/10.1172/JCI138003>
42. (U) Casanova, L. M.; Jeon, S.; Rutala, W. A.; Weber, D. J.; Sobsey, M. D., Effects of air temperature and relative humidity on coronavirus survival on surfaces. *Applied and environmental microbiology* **2010**, *76* (9), 2712-2717. <https://www.ncbi.nlm.nih.gov/pubmed/20228108>
43. (U) CDC, 2019 Novel Coronavirus RT-PCR Identification Protocols.
<https://www.cdc.gov/coronavirus/2019-ncov/lab/rt-pcr-detection-instructions.html>.
44. (U) CDC, Confirmed 2019-nCoV Cases Globally. <https://www.cdc.gov/coronavirus/2019-ncov/locations-confirmed-cases.html>.
45. (U) CDC, Interim Clinical Guidance for Management of Patients with Confirmed Coronavirus Disease 2019 (COVID-19). <https://www.cdc.gov/coronavirus/2019-ncov/hcp/clinical-guidance-management-patients.html>.
46. (U) CDC, Interim healthcare infection prevention and control recommendations for patients under investigation for 2019 novel coronavirus. <https://www.cdc.gov/coronavirus/2019-ncov/infection-control.html>.

47. (U) CDC, Recommendation Regarding the Use of Cloth Face Coverings, Especially in Areas of Significant Community-Based Transmission. **2020**. <https://www.cdc.gov/coronavirus/2019-ncov/prevent-getting-sick/cloth-face-cover.html>
48. (U) CDC, Situation summary. <https://www.cdc.gov/coronavirus/2019-nCoV/summary.html>.
49. (U) CDC, Symptoms. <https://www.cdc.gov/coronavirus/2019-ncov/about/symptoms.html>.
50. (U) CDC, C., China's CDC detects a large number of new coronaviruses in the South China seafood market in Wuhan http://www.chinacdc.cn/yw_9324/202001/t20200127_211469.html (accessed 01/27/2020).
51. (U) CenterWatch, SAB Biotherapeutics wins BARDA MERS treatment contract. <https://www.centerwatch.com/articles/14742>.
52. (U) Chan, J. F.-W.; Yuan, S.; Kok, K.-H.; To, K. K.-W.; Chu, H.; Yang, J.; Xing, F.; Liu, J.; Yip, C. C.-Y.; Poon, R. W.-S.; Tsoi, H.-W.; Lo, S. K.-F.; Chan, K.-H.; Poon, V. K.-M.; Chan, W.-M.; Ip, J. D.; Cai, J.-P.; Cheng, V. C.-C.; Chen, H.; Hui, C. K.-M.; Yuen, K.-Y., A familial cluster of pneumonia associated with the 2019 novel coronavirus indicating person-to-person transmission: a study of a family cluster. *The Lancet* **2020**. <https://www.sciencedirect.com/science/article/pii/S0140673620301549>
53. (U) Chan, J. F.; Zhang, A. J.; Yuan, S.; Poon, V. K.; Chan, C. C.; Lee, A. C.; Chan, W. M.; Fan, Z.; Tsoi, H. W.; Wen, L.; Liang, R.; Cao, J.; Chen, Y.; Tang, K.; Luo, C.; Cai, J. P.; Kok, K. H.; Chu, H.; Chan, K. H.; Sridhar, S.; Chen, Z.; Chen, H.; To, K. K.; Yuen, K. Y., Simulation of the clinical and pathological manifestations of Coronavirus Disease 2019 (COVID-19) in golden Syrian hamster model: implications for disease pathogenesis and transmissibility. *Clin Infect Dis* **2020**. <https://www.ncbi.nlm.nih.gov/pubmed/32215622>
54. (U) Chan, K. H.; Peiris, J. S.; Lam, S. Y.; Poon, L. L.; Yuen, K. Y.; Seto, W. H., The Effects of Temperature and Relative Humidity on the Viability of the SARS Coronavirus. *Adv Virol* **2011**, *2011*, 734690. <https://www.ncbi.nlm.nih.gov/pmc/articles/PMC3265313/pdf/AV2011-734690.pdf>
55. (U) Chang, D.; Lin, M.; Wei, L.; Xie, L.; Zhu, G.; Dela Cruz, C. S.; Sharma, L., Epidemiologic and Clinical Characteristics of Novel Coronavirus Infections Involving 13 Patients Outside Wuhan, China. *JAMA* **2020**. <https://doi.org/10.1001/jama.2020.1623>
https://jamanetwork.com/journals/jama/articlepdf/2761043/jama_chang_2020_id_200007.pdf
56. (U) Changzheng, L. J. L., Experts in the medical treatment team: Wuhan's unexplained viral pneumonia patients can be controlled more. <https://www.cn-healthcare.com/article/20200110/content-528579.html>.
57. (U) Chen, C.; Cao, M.; Peng, L.; Guo, X.; Yang, F.; Wu, W.; Chen, L.; Yang, Y.; Liu, Y.; Wang, F., Coronavirus Disease-19 Among Children Outside Wuhan, China. *SSRN* **2020**. https://papers.ssrn.com/sol3/papers.cfm?abstract_id=3546071
58. (U) Chen, C.; Huang, J.; Cheng, Z.; Wu, J.; Chen, S.; Zhang, Y.; Chen, B.; Lu, M.; Luo, Y.; Zhang, J.; Yin, P.; Wang, X., Favipiravir versus Arbidol for COVID-19: A Randomized Clinical Trial. *medRxiv* **2020**, 2020.03.17.20037432. <https://www.medrxiv.org/content/medrxiv/early/2020/03/20/2020.03.17.20037432.full.pdf>
59. (U) Chen, H.; Guo, J.; Wang, C.; Luo, F.; Yu, X.; Zhang, W.; Li, J.; Zhao, D.; Xu, D.; Gong, Q., Clinical characteristics and intrauterine vertical transmission potential of COVID-19 infection in nine pregnant women: a retrospective review of medical records. *The Lancet* **2020**, *395* (10226), 809-815.
60. (U) Chen, N.; Zhou, M.; Dong, X.; Qu, J.; Gong, F.; Han, Y.; Qiu, Y.; Wang, J.; Liu, Y.; Wei, Y.; Xia, J.; Yu, T.; Zhang, X.; Zhang, L., Epidemiological and clinical characteristics of 99 cases of 2019 novel coronavirus pneumonia in Wuhan, China: a descriptive study. *Lancet* **2020**. <https://www.ncbi.nlm.nih.gov/pubmed/32007143>

61. (U) Chen, Y.; Peng, H.; Wang, L.; Zhao, Y.; Zeng, L.; Gao, H.; Liu, Y., Infants Born to Mothers With a New Coronavirus (COVID-19). *Frontiers in Pediatrics* **2020**, *8* (104).
<https://www.frontiersin.org/article/10.3389/fped.2020.00104>
62. (U) Chin, A.; Chu, J.; Perera, M.; Hui, K.; Yen, H.-L.; Chan, M.; Peiris, M.; Poon, L., Stability of SARS-CoV-2 in different environmental conditions. *medRxiv* **2020**, 2020.03.15.20036673.
<https://www.medrxiv.org/content/medrxiv/early/2020/03/27/2020.03.15.20036673.full.pdf>
63. (U) Chin, A. W. H.; Chu, J. T. S.; Perera, M. R. A.; Hui, K. P. Y.; Yen, H.-L.; Chan, M. C. W.; Peiris, M.; Poon, L. L. M., Stability of SARS-CoV-2 in different environmental conditions. *The Lancet Microbe*.
[https://doi.org/10.1016/S2666-5247\(20\)30003-3](https://doi.org/10.1016/S2666-5247(20)30003-3)
64. (U) Chughtai, A. A.; Seale, H.; MacIntyre, C. R., Use of cloth masks in the practice of infection control—evidence and policy gaps. *Int J Infect Control* **2013**, *9* (3), doi: 10.3396/IJIC.v9i3.020.13.
65. (U) Ćirić, J., Is Iceland's coronavirus testing showing that 50% of cases have no symptoms? *Iceland Review* 02 April, 2020.
66. (U) Coalition, C.-C. R., Global coalition to accelerate COVID-19 clinical research in resource-limited settings. *The Lancet* **2020**. <http://www.sciencedirect.com/science/article/pii/S0140673620307984>
67. (U) Cockrell, A. S.; Yount, B. L.; Scobey, T.; Jensen, K.; Douglas, M.; Beall, A.; Tang, X.-C.; Marasco, W. A.; Heise, M. T.; Baric, R. S., A mouse model for MERS coronavirus-induced acute respiratory distress syndrome. *Nature microbiology* **2016**, *2* (2), 1-11.
68. (U) Cohen, J., Mining coronavirus genomes for clues to the outbreak's origins. *Science* **2020**.
69. (U) Cohen, J., Vaccine designers take first shots at COVID-19. *AAAS* **2020**, *368* (6486), 14-16.
<https://science.sciencemag.org/content/368/6486/14>
70. (U) Cohen, J., Wuhan seafood market may not be source of novel virus spreading globally.
<https://www.sciencemag.org/news/2020/01/wuhan-seafood-market-may-not-be-source-novel-virus-spreading-globally> (accessed 01/27/2020).
71. (U) Control, E. E. C. f. D. P. a., *Interim guidance for environmental cleaning in non-healthcare facilities exposed to SARS-CoV-2*; European Centre for Disease Prevention and Control: European Centre for Disease Prevention and Control, 2020. <https://www.ecdc.europa.eu/en/publications-data/interim-guidance-environmental-cleaning-non-healthcare-facilities-exposed-2019#no-link>
72. (U) Corman, V. M.; Landt, O.; Kaiser, M.; Molenkamp, R.; Meijer, A.; Chu, D. K.; Bleicker, T.; Brunink, S.; Schneider, J.; Schmidt, M. L.; Mulders, D. G.; Haagmans, B. L.; van der Veer, B.; van den Brink, S.; Wijsman, L.; Goderski, G.; Romette, J. L.; Ellis, J.; Zambon, M.; Peiris, M.; Goossens, H.; Reusken, C.; Koopmans, M. P.; Drosten, C., Detection of 2019 novel coronavirus (2019-nCoV) by real-time RT-PCR. *Euro Surveill* **2020**, *25* (3). <https://www.ncbi.nlm.nih.gov/pubmed/31992387>
73. (U) Correction, O. D. o. R. a., COVID-19 Inmate Testing, Updated 4/20/2020. 2020.
<https://drc.ohio.gov/Portals/0/DRC%20COVID-19%20Information%2004-20-2020%20%201304.pdf>
74. (U) Coughlin, M. M.; Prabhakar, B. S., Neutralizing human monoclonal antibodies to severe acute respiratory syndrome coronavirus: target, mechanism of action, and therapeutic potential. *Reviews in medical virology* **2012**, *22* (1), 2-17. <https://www.ncbi.nlm.nih.gov/pubmed/21905149>
75. (U) Cowling, B. J.; Ali, S. T.; Ng, T. W. Y.; Tsang, T. K.; Li, J. C. M.; Fong, M. W.; Liao, Q.; Kwan, M. Y.; Lee, S. L.; Chiu, S. S.; Wu, J. T.; Wu, P.; Leung, G. M., Impact assessment of non-pharmaceutical interventions against COVID-19 and influenza in Hong Kong: an observational study. *medRxiv* **2020**, 2020.03.12.20034660.
<https://www.medrxiv.org/content/medrxiv/early/2020/03/16/2020.03.12.20034660.full.pdf>
76. (U) Daily, H., Wuhan Institute of Virology, Chinese Academy of Sciences and others have found that 3 drugs have a good inhibitory effect on new coronavirus. Chen, L., Ed. 2020.
http://news.cnhubei.com/content/2020-01/28/content_12656365.html
77. (U) Dandekar, R.; Barbastathis, G., Neural Network aided quarantine control model estimation of global Covid-19 spread. *arXiv preprint arXiv:2004.02752* **2020**.

78. (U) Dato, V. M.; Hostler, D.; Hahn, M. E., Simple respiratory mask. *Emerg Infect Dis* **2006**, *12* (6), 1033-4. <https://www.ncbi.nlm.nih.gov/pubmed/16752475>
79. (U) Davies, A.; Thompson, K. A.; Giri, K.; Kafatos, G.; Walker, J.; Bennett, A., Testing the efficacy of homemade masks: would they protect in an influenza pandemic? *Disaster Med Public Health Prep* **2013**, *7* (4), 413-8. <https://www.ncbi.nlm.nih.gov/pubmed/24229526>
80. (U) De Albuquerque, N.; Baig, E.; Ma, X.; Zhang, J.; He, W.; Rowe, A.; Habal, M.; Liu, M.; Shalev, I.; Downey, G. P.; Gorczynski, R.; Butany, J.; Leibowitz, J.; Weiss, S. R.; McGilvray, I. D.; Phillips, M. J.; Fish, E. N.; Levy, G. A., Murine hepatitis virus strain 1 produces a clinically relevant model of severe acute respiratory syndrome in A/J mice. *J Virol* **2006**, *80* (21), 10382-94. <https://www.ncbi.nlm.nih.gov/pmc/articles/PMC1641767/pdf/0747-06.pdf>
81. (U) de Haan, C. A. M.; Haijema, B. J.; Schellen, P.; Schreur, P. W.; te Lintel, E.; Vennema, H.; Rottier, P. J. M., Cleavage of Group 1 Coronavirus Spike Proteins: How Furin Cleavage Is Traded Off against Heparan Sulfate Binding upon Cell Culture Adaptation. *Journal of Virology* **2008**, *82* (12), 6078-6083. <https://jvi.asm.org/content/jvi/82/12/6078.full.pdf>
82. (U) Dediego, M. L.; Pewe, L.; Alvarez, E.; Rejas, M. T.; Perlman, S.; Enjuanes, L., Pathogenicity of severe acute respiratory coronavirus deletion mutants in hACE-2 transgenic mice. *Virology* **2008**, *376* (2), 379-389. <https://www.ncbi.nlm.nih.gov/pmc/articles/PMC2810402/>
83. (U) Deng, W.; Bao, L.; Gao, H.; Xiang, Z.; Qu, Y.; Song, Z.; Gong, S.; Liu, J.; Liu, J.; Yu, P.; Qi, F.; Xu, Y.; Li, F.; Xiao, C.; Lv, Q.; Xue, J.; Wei, Q.; Liu, M.; Wang, G.; Wang, S.; Yu, H.; Liu, X.; Zhao, W.; Han, Y.; Qin, C., Ocular conjunctival inoculation of SARS-CoV-2 can cause mild COVID-19 in Rhesus macaques. *bioRxiv* **2020**.
84. (U) Dong, N.; Yang, X.; Ye, L.; Chen, K.; Chan, E. W.-C.; Yang, M.; Chen, S., Genomic and protein structure modelling analysis depicts the origin and infectivity of 2019-nCoV, a new coronavirus which caused a pneumonia outbreak in Wuhan, China. *bioRxiv* **2020**, 2020.01.20.913368. <https://www.biorxiv.org/content/biorxiv/early/2020/01/22/2020.01.20.913368.full.pdf>
85. (U) Dong, Y.; Mo, X.; Hu, Y.; Qi, X.; Jiang, F.; Jiang, Z.; Tong, S., Epidemiological Characteristics of 2143 Pediatric Patients With 2019 Coronavirus Disease in China. *Pediatrics* **2020**, e20200702. <https://pediatrics.aappublications.org/content/pediatrics/early/2020/03/16/peds.2020-0702.full.pdf>
86. (U) Du, Z.; Xu, X.; Wu, Y.; Wang, L.; Cowling, B. J.; Meyers, L. A., The serial interval of COVID-19 from publicly reported confirmed cases. *medRxiv* **2020**, 2020.02.19.20025452. <https://www.medrxiv.org/content/medrxiv/early/2020/03/13/2020.02.19.20025452.full.pdf>
87. (U) Duan, S.; Zhao, X.; Wen, R.; Huang, J.-j.; Pi, G.; Zhang, S.; Han, J.; Bi, S.; Ruan, L.; Dong, X.-p., Stability of SARS coronavirus in human specimens and environment and its sensitivity to heating and UV irradiation. *Biomedical and environmental sciences: BES* **2003**, *16* (3), 246-255.
88. (U) Duan, S. M.; Zhao, X. S.; Wen, R. F.; Huang, J. J.; Pi, G. H.; Zhang, S. X.; Han, J.; Bi, S. L.; Ruan, L.; Dong, X. P., Stability of SARS coronavirus in human specimens and environment and its sensitivity to heating and UV irradiation. *Biomed Environ Sci* **2003**, *16* (3), 246-55.
89. (U) EuroTimes, Pfizer/BioNTech target April vaccine trial launch. *EuroTimes* 2020.
90. (U) FDA, *Emergency Use Authorization*; Food and Drug Administration: 2020. <https://www.fda.gov/media/136529/download>
91. (U) FDA, FAQs on Shortages of Surgical Masks and Gowns. <https://www.fda.gov/medical-devices/personal-protective-equipment-infection-control/faqs-shortages-surgical-masks-and-gowns#kn95>.
92. (U) FDA, *ID NOW COVID-19*; Food and Drug Administration: 2020. <https://www.fda.gov/media/136525/download>
93. (U) FDA, *Investigational COVID-19 Convalescent Plasma - Emergency INDs*; Food and Drug Administration: 2020. <https://www.fda.gov/vaccines-blood-biologics/investigational-new-drug-ind-or-device-exemption-ide-process-cber/investigational-covid-19-convalescent-plasma-emergency-inds>

Updated 4/21/2020

94. (U) FDA, Policy for Diagnostics Testing in Laboratories Certified to Perform High Complexity Testing under CLIA prior to Emergency Use Authorization for Coronavirus Disease-2019 during the Public Health Emergency; Immediately in Effect Guidance for Industry and Food and Drug Administration Staff. 2020. <https://www.regulations.gov/docket?D=FDA-2020-D-0987>
95. (U) Ferguson, N.; Laydon, D.; Nedjati-Gilani, G.; Imai, N.; Ainslie, K.; Baguelin, M.; Bhatia, S.; Boonyasiri, A.; Cucunuba, Z.; Cuomo-Dannenburg, G.; Dighe, A.; Dorigatti, I.; Fu, H.; Gaythorpe, K.; Green, W.; Hamlet, A.; Hinsley, W.; Okell, L.; van Elsland, S.; Thompson, H.; Verity, R.; Volz, E.; Wang, H.; Wang, Y.; Walker, P.; Walters, C.; Winskill, P.; Whittaker, C.; Donnelly, C.; Riley, S.; Ghani, A., *Impact of non-pharmaceutical interventions (NPIs) to reduce COVID-19 mortality and healthcare demand*; 2020. <https://www.imperial.ac.uk/media/imperial-college/medicine/sph/ide/gida-fellowships/Imperial-College-COVID19-NPI-modelling-16-03-2020.pdf>
96. (U) Feuerstein, A.; Herper, M., Early peek at data on Gilead coronavirus drug suggests patients are responding to treatment. *Stat* 2020.
97. (U) Garg, S., Hospitalization Rates and Characteristics of Patients Hospitalized with Laboratory-Confirmed Coronavirus Disease 2019—COVID-NET, 14 States, March 1–30, 2020. *MMWR. Morbidity and Mortality Weekly Report* 2020, 69.
98. (U) Gautier, J.-F.; Ravussin, Y., A New Symptom of COVID-19: Loss of Taste and Smell. *Obesity* 2020, n/a (n/a). <https://doi.org/10.1002/oby.22809>
99. (U) Gautret, P.; Lagier, J.-C.; Parola, P.; Meddeb, L.; Mailhe, M.; Doudier, B.; Courjon, J.; Giordanengo, V.; Vieira, V. E.; Dupont, H. T., Hydroxychloroquine and azithromycin as a treatment of COVID-19: results of an open-label non-randomized clinical trial. *International Journal of Antimicrobial Agents* 2020, 105949.
100. (U) GitHub Inc., Reproducible analyses for rejecting rare genomic inversions in SARS-CoV-2. https://github.com/alexcritschroph/sars_cov_2_inversion (accessed 04 April).
101. (U) Goyal, P.; Choi, J. J.; Pinheiro, L. C.; Schenck, E. J.; Chen, R.; Jabri, A.; Satlin, M. J.; Campion, T. R.; Nahid, M.; Ringel, J. B.; Hoffman, K. L.; Alshak, M. N.; Li, H. A.; Wehmeyer, G. T.; Rajan, M.; Reshetnyak, E.; Hupert, N.; Horn, E. M.; Martinez, F. J.; Gulick, R. M.; Safford, M. M., Clinical Characteristics of Covid-19 in New York City. *New England Journal of Medicine* 2020. <https://www.nejm.org/doi/full/10.1056/NEJMc2010419>
102. (U) Grein, J.; Ohmagari, N.; Shin, D.; Diaz, G.; Asperges, E.; Castagna, A.; Feldt, T.; Green, G.; Green, M. L.; Lescure, F.-X.; Nicastri, E.; Oda, R.; Yo, K.; Quiros-Roldan, E.; Studemeister, A.; Redinski, J.; Ahmed, S.; Bennett, J.; Chelliah, D.; Chen, D.; Chihara, S.; Cohen, S. H.; Cunningham, J.; D'Arminio Monforte, A.; Ismail, S.; Kato, H.; Lapadula, G.; L'Her, E.; Maeno, T.; Majumder, S.; Massari, M.; Mora-Rillo, M.; Mutoh, Y.; Nguyen, D.; Verweij, E.; Zoufaly, A.; Osinusi, A. O.; DeZure, A.; Zhao, Y.; Zhong, L.; Chokkalingam, A.; Elboudwarej, E.; Telep, L.; Timbs, L.; Henne, I.; Sellers, S.; Cao, H.; Tan, S. K.; Winterbourne, L.; Desai, P.; Mera, R.; Gaggari, A.; Myers, R. P.; Brainard, D. M.; Childs, R.; Flanigan, T., Compassionate Use of Remdesivir for Patients with Severe Covid-19. *New England Journal of Medicine* 2020. <https://www.nejm.org/doi/full/10.1056/NEJMoa2007016>
103. (U) Guan, L.; Zhou, L.; Zhang, J.; Peng, W.; Chen, R., More awareness is needed for severe acute respiratory syndrome coronavirus 2019 transmission through exhaled air during non-invasive respiratory support: experience from China. *European Respiratory Journal* 2020, 55 (3), 2000352. <https://erj.ersjournals.com/content/erj/55/3/2000352.full.pdf>
104. (U) Guan, W.-j.; Ni, Z.-y.; Hu, Y.; Liang, W.-h.; Ou, C.-q.; He, J.-x.; Liu, L.; Shan, H.; Lei, C.-l.; Hui, D. S.; Du, B.; Li, L.-j.; Zeng, G.; Yuen, K.-Y.; Chen, R.-c.; Tang, C.-l.; Wang, T.; Chen, P.-y.; Xiang, J.; Li, S.-y.; Wang, J.-l.; Liang, Z.-j.; Peng, Y.-x.; Wei, L.; Liu, Y.; Hu, Y.-h.; Peng, P.; Wang, J.-m.; Liu, J.-y.; Chen, Z.; Li, G.; Zheng, Z.-j.; Qiu, S.-q.; Luo, J.; Ye, C.-j.; Zhu, S.-y.; Zhong, N.-s., Clinical characteristics of 2019 novel coronavirus infection in China. *medRxiv* 2020, 2020.02.06.20020974. <https://www.medrxiv.org/content/medrxiv/early/2020/02/09/2020.02.06.20020974.full.pdf>

Updated 4/21/2020

105. (U) Guérin, C.; Reignier, J.; Richard, J.-C.; Beuret, P.; Gacouin, A.; Boulain, T.; Mercier, E.; Badet, M.; Mercat, A.; Baudin, O.; Clavel, M.; Chatellier, D.; Jaber, S.; Rosselli, S.; Mancebo, J.; Sirodot, M.; Hilbert, G.; Bengler, C.; Richecoeur, J.; Gainnier, M.; Bayle, F.; Bourdin, G.; Leray, V.; Girard, R.; Baboi, L.; Ayzac, L., Prone Positioning in Severe Acute Respiratory Distress Syndrome. *New England Journal of Medicine* **2013**, *368* (23), 2159-2168. <https://www.nejm.org/doi/full/10.1056/NEJMoa1214103>
106. (U) Guo, Z.; Wang, Z.; Zhang, S.; Li, X.; Li, L.; Li, C.; Cui, Y.; Fu, R.; Dong, Y.; Chi, X., Aerosol and Surface Distribution of Severe Acute Respiratory Syndrome Coronavirus 2 in Hospital Wards, Wuhan, China, 2020. *Emerging infectious diseases* **2020**, *26* (7).
107. (U) He, X.; Lau, E. H. Y.; Wu, P.; Deng, X.; Wang, J.; Hao, X.; Lau, Y. C.; Wong, J. Y.; Guan, Y.; Tan, X.; Mo, X.; Chen, Y.; Liao, B.; Chen, W.; Hu, F.; Zhang, Q.; Zhong, M.; Wu, Y.; Zhao, L.; Zhang, F.; Cowling, B. J.; Li, F.; Leung, G. M., Temporal dynamics in viral shedding and transmissibility of COVID-19. *Nature Medicine* **2020**. <https://doi.org/10.1038/s41591-020-0869-5>
108. (U) Herold, T.; Jurinovic, V.; Arnreich, C.; Hellmuth, J. C.; Bergwelt-Baildon, M.; Klein, M.; Weinberger, T., Level of IL-6 predicts respiratory failure in hospitalized symptomatic COVID-19 patients. *medRxiv* **2020**, 2020.04.01.20047381. <https://www.medrxiv.org/content/medrxiv/early/2020/04/04/2020.04.01.20047381.full.pdf>
109. (U) HHS, 2019-nCoV Update. 2020. https://www.hhs.gov/live/live-2/index.html?CDC_AA_refVal=https%3A%2F%2Fwww.cdc.gov%2Fmedia%2Fpress%2F2020%2Fa0128-hhs-coronavirus-update.html#11465
110. (U) Hill, V.; Rambaut, A., Phylodynamic analysis of SARS-CoV-2 | Update 2020-03-06. *Virological*: 2020. <http://virological.org/t/phylodynamic-analysis-of-sars-cov-2-update-2020-03-06/420>
111. (U) Hinton, D., qSARS-CoV-2 IgG/IgM Rapid Test- Letter of Authorization. FDA, Ed. FDA: 2020. <https://www.fda.gov/media/136622/download>
112. (U) Hoehl, S.; Berger, A.; Kortenbusch, M.; Cinatl, J.; Bojkova, D.; Rabenau, H.; Behrens, P.; Böddinghaus, B.; Götsch, U.; Naujoks, F.; Neumann, P.; Schork, J.; Tiarks-Jungk, P.; Walczok, A.; Eickmann, M.; Vehreschild, M. J. G. T.; Kann, G.; Wolf, T.; Gottschalk, R.; Ciesek, S., Evidence of SARS-CoV-2 Infection in Returning Travelers from Wuhan, China. *New England Journal of Medicine* **2020**. <https://www.nejm.org/doi/full/10.1056/NEJMc2001899>
113. (U) Hu, Z.; Song, C.; Xu, C.; Jin, G.; Chen, Y.; Xu, X.; Ma, H.; Chen, W.; Lin, Y.; Zheng, Y.; Wang, J.; Hu, Z.; Yi, Y.; Shen, H., Clinical characteristics of 24 asymptomatic infections with COVID-19 screened among close contacts in Nanjing, China. *Science China Life Sciences* **2020**. <https://doi.org/10.1007/s11427-020-1661-4>
114. (U) Huang, C.; Wang, Y.; Li, X.; Ren, L.; Zhao, J.; Hu, Y.; Zhang, L.; Fan, G.; Xu, J.; Gu, X.; Cheng, Z.; Yu, T.; Xia, J.; Wei, Y.; Wu, W.; Xie, X.; Yin, W.; Li, H.; Liu, M.; Xiao, Y.; Gao, H.; Guo, L.; Xie, J.; Wang, G.; Jiang, R.; Gao, Z.; Jin, Q.; Wang, J.; Cao, B., Clinical features of patients infected with 2019 novel coronavirus in Wuhan, China. *The Lancet* **2020**. [https://www.thelancet.com/journals/lancet/article/PIIS0140-6736\(20\)30183-5/fulltext](https://www.thelancet.com/journals/lancet/article/PIIS0140-6736(20)30183-5/fulltext)
115. (U) Hulkower, R. L.; Casanova, L. M.; Rutala, W. A.; Weber, D. J.; Sobsey, M. D., Inactivation of surrogate coronaviruses on hard surfaces by health care germicides. *American journal of infection control* **2011**, *39* (5), 401-407. <https://www.sciencedirect.com/science/article/pii/S0196655310009004>
116. (U) IDEXX, Leading Veterinary Diagnostic Company Sees No COVID-19 Cases in Pets. IDEXX: 2020. <https://www.idexx.com/en/about-idexx/news/no-covid-19-cases-pets/>
117. (U) Iwata, K.; Doi, A.; Miyakoshi, C., Was School Closure Effective in Mitigating Coronavirus Disease 2019 (COVID-19)? Time Series Analysis Using Bayesian Inference. **2020**.
118. (U) JHU, Coronavirus COVID-19 Global Cases by Johns Hopkins CSSE. <https://gisanddata.maps.arcgis.com/apps/opsdashboard/index.html#/bda7594740fd40299423467b48e9ecf6>.

119. (U) Jing, C.; Wenjie, S.; Jianping, H.; Michelle, G.; Jing, W.; Guiqing, H., Indirect Virus Transmission in Cluster of COVID-19 Cases, Wenzhou, China, 2020. *Emerging Infectious Disease journal* **2020**, *26* (6). https://wwwnc.cdc.gov/eid/article/26/6/20-0412_article
120. (U) Johndrow, J. E.; Lum, K.; Ball, P., Estimating SARS-CoV-2-positive Americans using deaths-only data. *arXiv preprint arXiv:2004.02605* **2020**.
121. (U) Johnson, J. a., Johnson & Johnson Announces a Lead Vaccine Candidate for COVID-19; Landmark New Partnership with U.S. Department of Health & Human Services; and Commitment to Supply One Billion Vaccines Worldwide for Emergency Pandemic Use. Johnson and Johnson: 2020. <https://www.jnj.com/johnson-johnson-announces-a-lead-vaccine-candidate-for-covid-19-landmark-new-partnership-with-u-s-department-of-health-human-services-and-commitment-to-supply-one-billion-vaccines-worldwide-for-emergency-pandemic-use>
122. (U) Joseph, A., CDC developing serologic tests that could reveal full scope of U.S. coronavirus outbreak. *STAT* **2020**.
123. (U) Karamitros, T.; Papadopoulou, G.; Bousali, M.; Mexias, A.; Tsiodras, S.; Mentis, A., SARS-CoV-2 exhibits intra-host genomic plasticity and low-frequency polymorphic quasispecies. *bioRxiv* **2020**, 2020.03.27.009480. <http://biorxiv.org/content/early/2020/03/28/2020.03.27.009480.abstract>
124. (U) Kim, Y.-I.; Kim, S.-G.; Kim, S.-M.; Kim, E.-H.; Park, S.-J.; Yu, K.-M.; Chang, J.-H.; Kim, E. J.; Lee, S.; Casel, M. A. B.; Um, J.; Song, M.-S.; Jeong, H. W.; Lai, V. D.; Kim, Y.; Chin, B. S.; Park, J.-S.; Chung, K.-H.; Foo, S.-S.; Poo, H.; Mo, I.-P.; Lee, O.-J.; Webby, R. J.; Jung, J. U.; Choi, Y. K., Infection and Rapid Transmission of SARS-CoV-2 in Ferrets. *Cell Host & Microbe* **2020**. <http://www.sciencedirect.com/science/article/pii/S1931312820301876>
125. (U) Kissler, S. M.; Tedijanto, C.; Goldstein, E.; Grad, Y. H.; Lipsitch, M., Projecting the transmission dynamics of SARS-CoV-2 through the postpandemic period. *Science* **2020**, eabb5793. <https://science.sciencemag.org/content/sci/early/2020/04/14/science.abb5793.full.pdf>
126. (U) Klok, F.; Kruip, M.; van der Meer, N.; Arbous, M.; Gommers, D.; Kant, K.; Kaptein, F.; van Paassen, J.; Stals, M.; Huisman, M., Incidence of thrombotic complications in critically ill ICU patients with COVID-19. *Thrombosis Research* **2020**.
127. (U) Kraemer, M. U. G.; Yang, C.-H.; Gutierrez, B.; Wu, C.-H.; Klein, B.; Pigott, D. M.; du Plessis, L.; Faria, N. R.; Li, R.; Hanage, W. P.; Brownstein, J. S.; Layan, M.; Vespignani, A.; Tian, H.; Dye, C.; Pybus, O. G.; Scarpino, S. V., The effect of human mobility and control measures on the COVID-19 epidemic in China. *Science* **2020**, eabb4218. <https://science.sciencemag.org/content/sci/early/2020/03/25/science.abb4218.full.pdf>
128. (U) Krantz, S. G.; Rao, A. S. S., Level of under-reporting including under-diagnosis before the first peak of COVID-19 in various countries: Preliminary Retrospective Results Based on Wavelets and Deterministic Modeling. *Infection Control & Hospital Epidemiology* **2020**, 1-8.
129. (U) Kratzel, A.; Todt, D.; V'kovski, P.; Steiner, S.; Gultom, M. L.; Thao, T. T. N.; Ebert, N.; Holwerda, M.; Steinmann, J.; Niemeyer, D.; Dijkman, R.; Kampf, G.; Drosten, C.; Steinmann, E.; Thiel, V.; Pfaender, S., Efficient inactivation of SARS-CoV-2 by WHO-recommended hand rub formulations and alcohols. *bioRxiv* **2020**, 2020.03.10.986711. <https://www.biorxiv.org/content/biorxiv/early/2020/03/17/2020.03.10.986711.full.pdf>
130. (U) Kupferschmidt, K.; Cohen, J., WHO launches global megatrial of the four most promising coronavirus treatments. *Science* **2020**.
131. (U) Lai, M. Y.; Cheng, P. K.; Lim, W. W., Survival of severe acute respiratory syndrome coronavirus. *Clinical Infectious Diseases* **2005**, *41* (7), e67-e71. <https://academic.oup.com/cid/article/41/7/e67/310340>
132. (U) Lam, T. T.-Y.; Shum, M. H.-H.; Zhu, H.-C.; Tong, Y.-G.; Ni, X.-B.; Liao, Y.-S.; Wei, W.; Cheung, W. Y.-M.; Li, W.-J.; Li, L.-F.; Leung, G. M.; Holmes, E. C.; Hu, Y.-L.; Guan, Y., Identifying SARS-CoV-2 related coronaviruses in Malayan pangolins. *Nature* **2020**. <https://doi.org/10.1038/s41586-020-2169-0>

133. (U) Lan, L.; Xu, D.; Ye, G.; Xia, C.; Wang, S.; Li, Y.; Xu, H., Positive RT-PCR Test Results in Patients Recovered From COVID-19. *Jama* **2020**. <https://jamanetwork.com/journals/jama/fullarticle/2762452>
134. (U) Lasry, A.; Kidder, D.; Hast, M.; Poovey, J.; Sunshine, G.; Zviedrite, N.; Ahmed, F.; Ethier, K. A., Timing of community mitigation and changes in reported COVID-19 and community mobility—four US metropolitan areas, February 26–April 1, 2020. **2020**.
135. (U) Lau, S., Coronavirus: WHO official says there's no evidence of 'reinfected' patients in China <https://www.scmp.com/news/china/society/article/3074045/coronavirus-who-official-says-theres-no-evidence-reinfected>.
136. (U) Lauer, S. A.; Grantz, K. H.; Bi, Q.; Jones, F. K.; Zheng, Q.; Meredith, H. R.; Azman, A. S.; Reich, N. G.; Lessler, J., The Incubation Period of Coronavirus Disease 2019 (COVID-19) From Publicly Reported Confirmed Cases: Estimation and Application. *Annals of Internal Medicine* **2020**. <https://doi.org/10.7326/M20-0504>
137. (U) Leung, K.; Wu, J. T.; Liu, D.; Leung, G. M., First-wave COVID-19 transmissibility and severity in China outside Hubei after control measures, and second-wave scenario planning: a modelling impact assessment. *The Lancet*. [https://doi.org/10.1016/S0140-6736\(20\)30746-7](https://doi.org/10.1016/S0140-6736(20)30746-7)
138. (U) Leung, N. H. L.; Chu, D. K. W.; Shiu, E. Y. C.; Chan, K.-H.; McDevitt, J. J.; Hau, B. J. P.; Yen, H.-L.; Li, Y.; Ip, D. K. M.; Peiris, J. S. M.; Seto, W.-H.; Leung, G. M.; Milton, D. K.; Cowling, B. J., Respiratory virus shedding in exhaled breath and efficacy of face masks. *Nature Medicine* **2020**. <https://doi.org/10.1038/s41591-020-0843-2>
139. (U) Levine, J., Scientists race to develop vaccine to deadly China coronavirus. <https://nypost.com/2020/01/25/scientists-race-to-develop-vaccine-to-deadly-china-coronavirus/>.
140. (U) Lewis, D., Is the coronavirus airborne? Experts can't agree. *Nature* **2020**. 10.1038/d41586-020-00974-w
141. (U) Li, K.; Wohlford-Lenane, C.; Perlman, S.; Zhao, J.; Jewell, A. K.; Reznikov, L. R.; Gibson-Corley, K. N.; Meyerholz, D. K.; McCray, P. B., Jr., Middle East Respiratory Syndrome Coronavirus Causes Multiple Organ Damage and Lethal Disease in Mice Transgenic for Human Dipeptidyl Peptidase 4. *J Infect Dis* **2016**, 213 (5), 712-22. <https://www.ncbi.nlm.nih.gov/pmc/articles/PMC4747621/pdf/jiv499.pdf>
142. (U) Li, Q.; Guan, X.; Wu, P.; Wang, X.; Zhou, L.; Tong, Y.; Ren, R.; Leung, K. S. M.; Lau, E. H. Y.; Wong, J. Y.; Xing, X.; Xiang, N.; Wu, Y.; Li, C.; Chen, Q.; Li, D.; Liu, T.; Zhao, J.; Liu, M.; Tu, W.; Chen, C.; Jin, L.; Yang, R.; Wang, Q.; Zhou, S.; Wang, R.; Liu, H.; Luo, Y.; Liu, Y.; Shao, G.; Li, H.; Tao, Z.; Yang, Y.; Deng, Z.; Liu, B.; Ma, Z.; Zhang, Y.; Shi, G.; Lam, T. T. Y.; Wu, J. T.; Gao, G. F.; Cowling, B. J.; Yang, B.; Leung, G. M.; Feng, Z., Early Transmission Dynamics in Wuhan, China, of Novel Coronavirus–Infected Pneumonia. *New England Journal of Medicine* **2020**. <https://www.nejm.org/doi/full/10.1056/NEJMoa2001316>
- <https://www.nejm.org/doi/10.1056/NEJMoa2001316>
143. (U) Li, R.; Pei, S.; Chen, B.; Song, Y.; Zhang, T.; Yang, W.; Shaman, J., Substantial undocumented infection facilitates the rapid dissemination of novel coronavirus (SARS-CoV2). *Science* **2020**, eabb3221. <https://science.sciencemag.org/content/sci/early/2020/03/13/science.abb3221.full.pdf>
144. (U) Li, W.; Zhang, B.; Lu, J.; Liu, S.; Chang, Z.; Cao, P.; Liu, X.; Zhang, P.; Ling, Y.; Tao, K.; Chen, J., The characteristics of household transmission of COVID-19. *Clinical Infectious Diseases* **2020**. <https://doi.org/10.1093/cid/ciaa450>
145. (U) Li, X.; Zai, J.; Zhao, Q.; Nie, Q.; Li, Y.; Foley, B. T.; Chaillon, A., Evolutionary history, potential intermediate animal host, and cross-species analyses of SARS-CoV-2. *Journal of Medical Virology* **2020**, n/a (n/a). <https://onlinelibrary.wiley.com/doi/abs/10.1002/jmv.25731>
146. (U) Liu, H.; Wang, L.-L.; Zhao, S.-J.; Kwak-Kim, J.; Mor, G.; Liao, A.-H., Why are pregnant women susceptible to viral infection: an immunological viewpoint? *Journal of Reproductive Immunology* **2020**, 103122. <http://www.sciencedirect.com/science/article/pii/S0165037820300437>

147. (U) Liu, P.; Chen, W.; Chen, J.-P., Viral Metagenomics Revealed Sendai Virus and Coronavirus Infection of Malayan Pangolins (*Manis javanica*). *Viruses* **2019**, *11* (11), 979.
<https://www.mdpi.com/1999-4915/11/11/979>
148. (U) Liu, P.; Jiang, J.-Z.; Wan, X.-F.; Hua, Y.; Wang, X.; Hou, F.; Chen, J.; Zou, J.; Chen, J., Are pangolins the intermediate host of the 2019 novel coronavirus (2019-nCoV) ? *bioRxiv* **2020**, 2020.02.18.954628.
<http://biorxiv.org/content/early/2020/02/20/2020.02.18.954628.abstract>
149. (U) Liu, W.; Zhang, Q.; Chen, J.; Xiang, R.; Song, H.; Shu, S.; Chen, L.; Liang, L.; Zhou, J.; You, L.; Wu, P.; Zhang, B.; Lu, Y.; Xia, L.; Huang, L.; Yang, Y.; Liu, F.; Semple, M. G.; Cowling, B. J.; Lan, K.; Sun, Z.; Yu, H.; Liu, Y., Detection of Covid-19 in Children in Early January 2020 in Wuhan, China. *New England Journal of Medicine* **2020**. <https://www.nejm.org/doi/full/10.1056/NEJMc2003717>
150. (U) Liu, Y.; Funk, S.; Flasche, S., *The Contribution of Pre-symptomatic Transmission to the COVID-19 Outbreak*; London School of Hygiene and Tropical Medicine: 2020.
<https://cmmid.github.io/topics/covid19/control-measures/pre-symptomatic-transmission.html>
151. (U) Liu, Y.; Ning, Z.; Chen, Y.; Guo, M.; Liu, Y.; Gali, N. K.; Sun, L.; Duan, Y.; Cai, J.; Westerdahl, D.; Liu, X.; Ho, K.-f.; Kan, H.; Fu, Q.; Lan, K., Aerodynamic Characteristics and RNA Concentration of SARS-CoV-2 Aerosol in Wuhan Hospitals during COVID-19 Outbreak. *bioRxiv* **2020**, 2020.03.08.982637.
<https://www.biorxiv.org/content/biorxiv/early/2020/03/10/2020.03.08.982637.full.pdf>
152. (U) Lowe, D., Hydroxychloroquine Update For April 6. (accessed April 7).
153. (U) Lu, J.; Plessis, L. d.; Liu, Z.; Hill, V.; Kang, M.; Lin, H.; Sun, J.; Francois, S.; Kraemer, M. U. G.; Faria, N. R.; McCrone, J. T.; Peng, J.; Xiong, Q.; Yuan, R.; Zeng, L.; Zhou, P.; Liang, C.; Yi, L.; Liu, J.; Xiao, J.; Hu, J.; Liu, T.; Ma, W.; Li, W.; Su, J.; Zheng, H.; Peng, B.; Fang, S.; Su, W.; Li, K.; Sun, R.; Bai, R.; Tang, X.; Liang, M.; Quick, J.; Song, T.; Rambaut, A.; Loman, N.; Raghwani, J.; Pybus, O.; Ke, C., Genomic epidemiology of SARS-CoV-2 in Guangdong Province, China. *medRxiv* **2020**, 2020.04.01.20047076.
<https://www.medrxiv.org/content/medrxiv/early/2020/04/04/2020.04.01.20047076.full.pdf>
154. (U) Lu, R.; Zhao, X.; Li, J.; Niu, P.; Yang, B.; Wu, H.; Wang, W.; Song, H.; Huang, B.; Zhu, N.; Bi, Y.; Ma, X.; Zhan, F.; Wang, L.; Hu, T.; Zhou, H.; Hu, Z.; Zhou, W.; Zhao, L.; Chen, J.; Meng, Y.; Wang, J.; Lin, Y.; Yuan, J.; Xie, Z.; Ma, J.; Liu, W. J.; Wang, D.; Xu, W.; Holmes, E. C.; Gao, G. F.; Wu, G.; Chen, W.; Shi, W.; Tan, W., Genomic characterisation and epidemiology of 2019 novel coronavirus: implications for virus origins and receptor binding. *The Lancet* **2020**. [https://doi.org/10.1016/S0140-6736\(20\)30251-8](https://doi.org/10.1016/S0140-6736(20)30251-8)
155. (U) Lu, X.; Zhang, L.; Du, H.; Zhang, J.; Li, Y. Y.; Qu, J.; Zhang, W.; Wang, Y.; Bao, S.; Li, Y.; Wu, C.; Liu, H.; Liu, D.; Shao, J.; Peng, X.; Yang, Y.; Liu, Z.; Xiang, Y.; Zhang, F.; Silva, R. M.; Pinkerton, K. E.; Shen, K.; Xiao, H.; Xu, S.; Wong, G. W. K., SARS-CoV-2 Infection in Children. *New England Journal of Medicine* **2020**. <https://www.nejm.org/doi/full/10.1056/NEJMc2005073>
156. (U) Luo, L.; Liu, D.; Liao, X.-l.; Wu, X.-b.; Jing, Q.-l.; Zheng, J.-z.; Liu, F.-h.; Yang, S.-g.; Bi, B.; Li, Z.-h.; Liu, J.-p.; Song, W.-q.; Zhu, W.; Wang, Z.-h.; Zhang, X.-r.; Chen, P.-l.; Liu, H.-m.; Cheng, X.; Cai, M.-c.; Huang, Q.-m.; Yang, P.; Yang, X.-f.; Huang, Z.-g.; Tang, J.-l.; Ma, Y.; Mao, C., Modes of contact and risk of transmission in COVID-19 among close contacts. *medRxiv* **2020**, 2020.03.24.20042606.
<https://www.medrxiv.org/content/medrxiv/early/2020/03/26/2020.03.24.20042606.full.pdf>
157. (U) Luo, W.; Majumder, M. S.; Liu, D.; Poirier, C.; Mandl, K. D.; Lipsitch, M.; Santillana, M., The role of absolute humidity on transmission rates of the COVID-19 outbreak. *medRxiv* **2020**, 2020.02.12.20022467.
<https://www.medrxiv.org/content/medrxiv/early/2020/02/17/2020.02.12.20022467.full.pdf>
158. (U) Mahevas, M.; Tran, V.-T.; Roumier, M.; Chabrol, A.; Paule, R.; Guillaud, C.; Gallien, S.; Lepeule, R.; Szwebel, T.-A.; Lescure, X.; Schlemmer, F.; Maignon, M.; Khellaf, M.; Crickx, E.; Terrier, B.; Morbieu, C.; Legendre, P.; Dang, J.; Schoindre, Y.; Pawlotski, J.-M.; Michel, M.; Perrodeau, E.; Carlier, N.; Roche, N.; De Lastours, V.; Mouthon, L.; Audureau, E.; Ravaud, P.; Godeau, B.; Costedoat, N., No evidence of clinical efficacy of hydroxychloroquine in patients hospitalized for COVID-19 infection with oxygen requirement: results of a study using routinely collected data to emulate a target trial. *medRxiv* **2020**,

2020.04.10.20060699.

<https://www.medrxiv.org/content/medrxiv/early/2020/04/14/2020.04.10.20060699.full.pdf>

159. (U) Majumder, M.; Mandl, K., Early transmissibility assessment of a novel coronavirus in Wuhan, China. *SSRN* **2020**. https://papers.ssrn.com/sol3/papers.cfm?abstract_id=3524675

160. (U) Malik, A. A.; Couzens, C.; Omer, S. B., COVID-19 related social distancing measures and reduction in city mobility. *medRxiv* **2020**, 2020.03.30.20048090.

<https://www.medrxiv.org/content/medrxiv/early/2020/04/06/2020.03.30.20048090.full.pdf>

161. (U) Mallapaty, S., Coronavirus can infect cats — dogs, not so much. *Nature* **2020**.

<https://www.nature.com/articles/d41586-020-00984-8>

162. (U) Mason, M., Hundreds of thousands in L.A. County may have been infected with coronavirus, study finds. *LA Times* 2020.

163. (U) Matthay, M. A.; Aldrich, J. M.; Gotts, J. E., Treatment for severe acute respiratory distress syndrome from COVID-19. *The Lancet Respiratory Medicine* **2020**. [https://doi.org/10.1016/S2213-2600\(20\)30127-2](https://doi.org/10.1016/S2213-2600(20)30127-2)

164. (U) Maxmen, A., How blood from coronavirus survivors might save lives. *Nature News* 2020.

165. (U) Melin, A. D.; Janiak, M. C.; Marrone, F.; Arora, P. S.; Higham, J. P., Comparative ACE2 variation and primate COVID-19 risk. *bioRxiv* **2020**, 2020.04.09.034967.

<https://www.biorxiv.org/content/biorxiv/early/2020/04/19/2020.04.09.034967.full.pdf>

166. (U) Menachery, V. D.; Dinnon, K. H.; Yount, B. L.; McAnarney, E. T.; Gralinski, L. E.; Hale, A.; Graham, R. L.; Scobey, T.; Anthony, S. J.; Wang, L.; Graham, B.; Randell, S. H.; Lipkin, W. I.; Baric, R. S., Trypsin Treatment Unlocks Barrier for Zoonotic Bat Coronavirus Infection. *Journal of Virology* **2020**, *94* (5), e01774-19. <https://jvi.asm.org/content/jvi/94/5/e01774-19.full.pdf>

167. (U) Mezher, M., FDA Grants First EUA for Blood Purification Device for COVID-19 Patients. *Regulatory Focus* 2020.

168. (U) Ministry for Foreign Affairs, Large scale testing of general population in Iceland underway In *Government of Iceland*, Government of Iceland: 2020.

<https://www.government.is/news/article/2020/03/15/Large-scale-testing-of-general-population-in-Iceland-underway/>

169. (U) Moriarty, L. F.; Plucinski, M. M.; Marston, B. J. e. a., Public Health Responses fo COVID-19 Outbreaks on Cruise Ships - Worldwide, February - March 2020. *MMWR* **2020**, (ePub: 23 March 2020).

<https://www.cdc.gov/mmwr/volumes/69/wr/mm6912e3.htm>

170. (U) Munster, V. J.; Feldmann, F.; Williamson, B. N.; van Doremalen, N.; Pérez-Pérez, L.; Schulz, J.; Meade-White, K.; Okumura, A.; Callison, J.; Brumbaugh, B.; Avanzato, V. A.; Rosenke, R.; Hanley, P. W.; Saturday, G.; Scott, D.; Fischer, E. R.; de Wit, E., Respiratory disease and virus shedding in rhesus macaques inoculated with SARS-CoV-2. *bioRxiv* **2020**, 2020.03.21.001628.

<https://www.biorxiv.org/content/biorxiv/early/2020/03/21/2020.03.21.001628.full.pdf>

171. (U) Muoio, D., Scanwell Health, myLAB Box unveil more at-home COVID-19 testing services. *MobiHealthNews* 20 March, 2020.

172. (U) Nadi, A., An at-home fingerprick blood test may help detect your exposure to coronavirus. *NBC NEWS* 04 April, 2020.

173. (U) NIH, NIH clinical trial of remdesivir to treat COVID-19 begins <https://www.nih.gov/news-events/news-releases/nih-clinical-trial-remdesivir-treat-covid-19-begins>.

174. (U) Okba, N.; Müller, M.; Li, W.; Wang, C.; GeurtsvanKessel, C.; Corman, V.; Lamers, M.; Sikkema, R.; de Bruin, E.; Chandler, F., Severe Acute Respiratory Syndrome Coronavirus 2-Specific Antibody Responses in Coronavirus Disease 2019 Patients. *Emerging infectious diseases* **2020**, *26* (7).

175. (U) Onder, G.; Rezza, G.; Brusaferro, S., Case-Fatality Rate and Characteristics of Patients Dying in Relation to COVID-19 in Italy. *JAMA* **2020**. <https://doi.org/10.1001/jama.2020.4683>

176. (U) Ong, S. W. X.; Tan, Y. K.; Chia, P. Y.; Lee, T. H.; Ng, O. T.; Wong, M. S. Y.; Marimuthu, K., Air, Surface Environmental, and Personal Protective Equipment Contamination by Severe Acute Respiratory Syndrome Coronavirus 2 (SARS-CoV-2) From a Symptomatic Patient. *Jama* **2020**.
https://jamanetwork.com/journals/jama/articlepdf/2762692/jama_ong_2020_id_200016.pdf
177. (U) Ortega, J. T.; Serrano, M. L.; Pujol, F. H.; Rangel, H. R., Role of changes in SARS-CoV-2 spike protein in the interaction with the human ACE2 receptor: An in silico analysis. *EXCLI journal* **2020**, *19*, 410-417. <https://pubmed.ncbi.nlm.nih.gov/32210742>
<https://www.ncbi.nlm.nih.gov/pmc/articles/PMC7081066/>
178. (U) Ou, J.; Zhou, Z.; Dai, R.; Zhang, J.; Lan, W.; Zhao, S.; Wu, J.; Seto, D.; Cui, L.; Zhang, G.; Zhang, Q., Emergence of RBD mutations in circulating SARS-CoV-2 strains enhancing the structural stability and human ACE2 receptor affinity of the spike protein. *bioRxiv* **2020**, 2020.03.15.991844.
<https://www.biorxiv.org/content/biorxiv/early/2020/04/20/2020.03.15.991844.full.pdf>
179. (U) Pan, A.; Liu, L.; Wang, C.; Guo, H.; Hao, X.; Wang, Q.; Huang, J.; He, N.; Yu, H.; Lin, X., Association of Public Health Interventions With the Epidemiology of the COVID-19 Outbreak in Wuhan, China. *JAMA* **2020**.
180. (U) Pan, F.; Ye, T.; Sun, P.; Gui, S.; Liang, B.; Li, L.; Zheng, D.; Wang, J.; Hesketh, R. L.; Yang, L.; Zheng, C., Time Course of Lung Changes On Chest CT During Recovery From 2019 Novel Coronavirus (COVID-19) Pneumonia. *Radiology* *0* (0), 200370. <https://pubs.rsna.org/doi/abs/10.1148/radiol.2020200370>
181. (U) Pan, L.; Mu, M.; Yang, P.; Sun, Y.; Wang, R.; Yan, J.; Li, P.; Hu, B.; Wang, J.; Hu, C.; Jin, Y.; Niu, X.; Ping, R.; Du, Y.; Li, T.; Xu, G.; Hu, Q.; Tu, L., Clinical characteristics of COVID-19 patients with digestive symptoms in Hubei, China: a descriptive, cross-sectional, multicenter study. *The American Journal of Gastroenterology* **2020**.
https://journals.lww.com/ajg/Documents/COVID_Digestive_Symptoms_AJG_Preproof.pdf
182. (U) Park, A., An At-Home Coronavirus Test May Be on the Way in the U.S. *TIME* 25 March, 2020.
183. (U) Park, S. W.; Champredon, D.; Earn, D. J. D.; Li, M.; Weitz, J. S.; Grenfell, B. T.; Dushoff, J., Reconciling early-outbreak preliminary estimates of the basic reproductive number and its uncertainty: a new framework and applications to the novel coronavirus (2019-nCoV) outbreak. **2020**, 1-13.
184. (U) Prem, K.; Liu, Y.; Russell, T. W.; Kucharski, A. J.; Eggo, R. M.; Davies, N.; Flasche, S.; Clifford, S.; Pearson, C. A. B.; Munday, J. D.; Abbott, S.; Gibbs, H.; Rosello, A.; Quilty, B. J.; Jombart, T.; Sun, F.; Diamond, C.; Gimma, A.; van Zandvoort, K.; Funk, S.; Jarvis, C. I.; Edmunds, W. J.; Bosse, N. I.; Hellewell, J.; Jit, M.; Klepac, P., The effect of control strategies to reduce social mixing on outcomes of the COVID-19 epidemic in Wuhan, China: a modelling study. *The Lancet Public Health* **2020**.
[https://doi.org/10.1016/S2468-2667\(20\)30073-6](https://doi.org/10.1016/S2468-2667(20)30073-6)
185. (U) Pyankov, O. V.; Bodnev, S. A.; Pyankova, O. G.; Agranovski, I. E., Survival of aerosolized coronavirus in the ambient air. *Journal of Aerosol Science* **2018**, *115*, 158-163.
<http://www.sciencedirect.com/science/article/pii/S0021850217302239>
186. (U) Qiu, H.; Wu, J.; Hong, L.; Luo, Y.; Song, Q.; Chen, D., Clinical and epidemiological features of 36 children with coronavirus disease 2019 (COVID-19) in Zhejiang, China: an observational cohort study. *The Lancet Infectious Diseases*. [https://doi.org/10.1016/S1473-3099\(20\)30198-5](https://doi.org/10.1016/S1473-3099(20)30198-5)
187. (U) Rabenau, H.; Kampf, G.; Cinatl, J.; Doerr, H., Efficacy of various disinfectants against SARS coronavirus. *Journal of Hospital Infection* **2005**, *61* (2), 107-111.
<https://www.sciencedirect.com/science/article/pii/S0195670105000447>
188. (U) Rabenau, H. F.; Cinatl, J.; Morgenstern, B.; Bauer, G.; Preiser, W.; Doerr, H. W., Stability and inactivation of SARS coronavirus. *Med Microbiol Immunol* **2005**, *194* (1-2), 1-6.
<https://link.springer.com/content/pdf/10.1007/s00430-004-0219-0.pdf>
189. (U) Rambaut, A., Phylodynamic analysis of nCoV-2019 genomes - 27-Jan-2020.
<http://virological.org/t/phylodynamic-analysis-of-ncov-2019-genomes-27-jan-2020/353>.

Updated 4/21/2020

190. (U) Rapid Expert Consultation, *Rapid Expert Consultation Update on SARS-CoV-2 Surface Stability and Incubation for the COVID-19 Pandemic (March 27, 2020)*. The National Academies Press: Washington, DC, 2020. <https://www.nap.edu/read/25763/chapter/1>
191. (U) Regalado, A., Blood tests show 14% of people are now immune to covid-19 in one town in Germany. *Technology Review* 2020.
192. (U) Remuzzi, A.; Remuzzi, G., COVID-19 and Italy: what next? *The Lancet* 2020. [https://doi.org/10.1016/S0140-6736\(20\)30627-9](https://doi.org/10.1016/S0140-6736(20)30627-9)
193. (U) Ren, X.; Liu, Y.; Chen, H.; Liu, W.; Guo, Z.; Chen, C.; Zhou, J.; Xiao, Q.; Jiang, G.-M.; Shan, H., Application and Optimization of RT-PCR in Diagnosis of SARS-CoV-2 Infection. *medRxiv* 2020.
194. (U) Rengasamy, S.; Eimer, B.; Shaffer, R. E., Simple respiratory protection--evaluation of the filtration performance of cloth masks and common fabric materials against 20-1000 nm size particles. *Ann Occup Hyg* 2010, 54 (7), 789-98. <https://www.ncbi.nlm.nih.gov/pubmed/20584862>
195. (U) Richard, M.; Kok, A.; de Meulder, D.; Bestebroer, T. M.; Lamers, M. M.; Okba, N. M. A.; Fentener van Vlissingen, M.; Rockx, B.; Haagmans, B. L.; Koopmans, M. P. G.; Fouchier, R. A. M.; Herfst, S., SARS-CoV-2 is transmitted via contact and via the air between ferrets. *bioRxiv* 2020, 2020.04.16.044503. <https://www.biorxiv.org/content/biorxiv/early/2020/04/17/2020.04.16.044503.full.pdf>
196. (U) Richter, W.; Hofacre, K.; Willenberg, Z., *Final Report for the Bioquell Hydrogen Peroxide Vapor (HPV) Decontamination for Reuse of N95 Respirators*; Battelle Memorial Institute: 2016. <http://wayback.archive-it.org/7993/20170113034232/http://www.fda.gov/downloads/EmergencyPreparedness/Counterterrorism/MedicalCountermeasures/MCMRegulatoryScience/UCM516998.pdf>
197. (U) Riou, J.; Althaus, C. L., Pattern of early human-to-human transmission of Wuhan 2019 novel coronavirus (2019-nCoV), December 2019 to January 2020. *Eurosurveillance* 2020, 25 (4), 2000058. <https://www.eurosurveillance.org/content/10.2807/1560-7917.ES.2020.25.4.2000058>
198. (U) Rivers, C.; Martin, E.; Watson, C.; Schoch-Spana, M.; Mullen, L.; Sell, T. K.; Gottlieb, S.; Warmbrod, K. L.; Hosangadi, D.; Kobokovich, A.; Potter, C.; Cicero, A.; Inglesby, T. V., *Public Health Principles for a Phased Reopening During COVID-19: Guidance for Governors*; Johns Hopkins Center for Health Security: 2020. https://www.centerforhealthsecurity.org/our-work/pubs_archive/pubs-pdfs/2020/reopening-guidance-governors.pdf
199. (U) Roberts, M., Coronavirus: US volunteers test first vaccine. *BBC* 2020.
200. (U) Robertson, D., nCoV's relationship to bat coronaviruses & recombination signals (no snakes) 2020. <http://virological.org/t/ncovs-relationship-to-bat-coronaviruses-recombination-signals-no-snakes/331>
201. (U) Rockx, B.; Kuiken, T.; Herfst, S.; Bestebroer, T.; Lamers, M. M.; Oude Munnink, B. B.; de Meulder, D.; van Amerongen, G.; van den Brand, J.; Okba, N. M. A.; Schipper, D.; van Run, P.; Leijten, L.; Sikkema, R.; Verschoor, E.; Verstrepen, B.; Bogers, W.; Langermans, J.; Drosten, C.; Fentener van Vlissingen, M.; Fouchier, R.; de Swart, R.; Koopmans, M.; Haagmans, B. L., Comparative pathogenesis of COVID-19, MERS, and SARS in a nonhuman primate model. *Science* 2020, eabb7314. <https://science.sciencemag.org/content/sci/early/2020/04/16/science.abb7314.full.pdf>
202. (U) Rothe, C.; Schunk, M.; Sothmann, P.; Bretzel, G.; Froeschl, G.; Wallrauch, C.; Zimmer, T.; Thiel, V.; Janke, C.; Guggemos, W.; Seilmaier, M.; Drosten, C.; Vollmar, P.; Zwirgmaier, K.; Zange, S.; Wölfel, R.; Hoelscher, M., Transmission of 2019-nCoV Infection from an Asymptomatic Contact in Germany. *New England Journal of Medicine* 2020. <https://www.nejm.org/doi/full/10.1056/NEJMc2001468>
<https://www.nejm.org/doi/10.1056/NEJMc2001468>
203. (U) Ruan, Q.; Yang, K.; Wang, W.; Jiang, L.; Song, J., Clinical predictors of mortality due to COVID-19 based on an analysis of data of 150 patients from Wuhan, China. *Intensive Care Medicine* 2020. <https://doi.org/10.1007/s00134-020-05991-x>

204. (U) Russell, T. W.; Hellewell, J.; Abbott, S.; Golding, N.; Gibbs, H.; Jarvis, C. I.; van Zandvoort, K.; group, C. n. w.; Flasche, S.; Eggo, R. M.; Edmunds, W. J.; Kucharski, A. J., Using a delay-adjusted case fatality ratio to estimate under-reporting. *CMMID*: 2020. https://cmmid.github.io/topics/covid19/severity/global_cfr_estimates.html
205. (U) Saknimit, M.; Inatsuki, I.; Sugiyama, Y.; Yagami, K., Virucidal efficacy of physico-chemical treatments against coronaviruses and parvoviruses of laboratory animals. *Jikken Dobutsu* **1988**, 37 (3), 341-5. https://www.istage.jst.go.jp/article/expanim1978/37/3/37_3_341/pdf
206. (U) Santarpia, J. L.; Rivera, D. N.; Herrera, V.; Morwitzer, M. J.; Creager, H.; Santarpia, G. W.; Crown, K. K.; Brett-Major, D.; Schnaubelt, E.; Broadhurst, M. J.; Lawler, J. V.; Reid, S. P.; Lowe, J. J., Transmission Potential of SARS-CoV-2 in Viral Shedding Observed at the University of Nebraska Medical Center. *medRxiv* **2020**, 2020.03.23.20039446. <https://www.medrxiv.org/content/medrxiv/early/2020/03/26/2020.03.23.20039446.1.full.pdf>
207. (U) Schnirring, L., New coronavirus infects health workers, spreads to Korea. <http://www.cidrap.umn.edu/news-perspective/2020/01/new-coronavirus-infects-health-workers-spreads-korea>.
208. (U) Schwartz, D. A., An analysis of 38 pregnant women with COVID-19, their newborn infants, and maternal-fetal transmission of SARS-CoV-2: maternal coronavirus infections and pregnancy outcomes. *Archives of Pathology & Laboratory Medicine* **2020**.
209. (U) Security, J. C. f. H., 2019-nCoV resources and updates on the emerging novel coronavirus. **2020**. <http://www.centerforhealthsecurity.org/resources/2019-nCoV/>
210. (U) Sheahan, T. P.; Sims, A. C.; Zhou, S.; Graham, R. L.; Pruijssers, A. J.; Agostini, M. L.; Leist, S. R.; Schäfer, A.; Dinno, K. H.; Stevens, L. J.; Chappell, J. D.; Lu, X.; Hughes, T. M.; George, A. S.; Hill, C. S.; Montgomery, S. A.; Brown, A. J.; Bluemling, G. R.; Natchus, M. G.; Saindane, M.; Kolykhalov, A. A.; Painter, G.; Harcourt, J.; Tamin, A.; Thornburg, N. J.; Swanstrom, R.; Denison, M. R.; Baric, R. S., An orally bioavailable broad-spectrum antiviral inhibits SARS-CoV-2 in human airway epithelial cell cultures and multiple coronaviruses in mice. *Science Translational Medicine* **2020**, eabb5883. <http://stm.sciencemag.org/content/early/2020/04/03/scitranslmed.abb5883.abstract>
211. (U) Shen, C.; Wang, Z.; Zhao, F.; Yang, Y.; Li, J.; Yuan, J.; Wang, F.; Li, D.; Yang, M.; Xing, L.; Wei, J.; Xiao, H.; Yang, Y.; Qu, J.; Qing, L.; Chen, L.; Xu, Z.; Peng, L.; Li, Y.; Zheng, H.; Chen, F.; Huang, K.; Jiang, Y.; Liu, D.; Zhang, Z.; Liu, Y.; Liu, L., Treatment of 5 Critically Ill Patients With COVID-19 With Convalescent Plasma. *JAMA* **2020**. <https://doi.org/10.1001/jama.2020.4783>
212. (U) Sheridan, C., Coronavirus and the race to distribute reliable diagnostics. <https://www.nature.com/articles/d41587-020-00002-2>.
213. (U) Shi, J.; Wen, Z.; Zhong, G.; Yang, H.; Wang, C.; Huang, B.; Liu, R.; He, X.; Shuai, L.; Sun, Z.; Zhao, Y.; Liu, P.; Liang, L.; Cui, P.; Wang, J.; Zhang, X.; Guan, Y.; Tan, W.; Wu, G.; Chen, H.; Bu, Z., Susceptibility of ferrets, cats, dogs, and other domesticated animals to SARS–coronavirus 2. *Science* **2020**, eabb7015. <https://science.sciencemag.org/content/sci/early/2020/04/07/science.abb7015.full.pdf>
214. (U) Shi, S.; Qin, M.; Shen, B.; Cai, Y.; Liu, T.; Yang, F.; Gong, W.; Liu, X.; Liang, J.; Zhao, Q.; Huang, H.; Yang, B.; Huang, C., Association of Cardiac Injury With Mortality in Hospitalized Patients With COVID-19 in Wuhan, China. *JAMA Cardiology* **2020**. <https://doi.org/10.1001/jamacardio.2020.0950>
215. (U) Su, H.; Yang, M.; Wan, C.; Yi, L.-X.; Tang, F.; Zhu, H.-Y.; Yi, F.; Yang, H.-C.; Fogo, A. B.; Nie, X.; Zhang, C., Renal histopathological analysis of 26 postmortem findings of patients with COVID-19 in China. *Kidney International*. <https://doi.org/10.1016/j.kint.2020.04.003>
216. (U) Su, Y. C.; Anderson, D. E.; Young, B. E.; Zhu, F.; Linster, M.; Kalimuddin, S.; Low, J. G.; Yan, Z.; Jayakumar, J.; Sun, L.; Yan, G. Z.; Mendenhall, I. H.; Leo, Y.-S.; Lye, D. C.; Wang, L.-F.; Smith, G. J., Discovery of a 382-nt deletion during the early evolution of SARS-CoV-2. *bioRxiv* **2020**, 2020.03.11.987222. <https://www.biorxiv.org/content/biorxiv/early/2020/03/12/2020.03.11.987222.full.pdf>

217. (U) ter Meulen, J.; van den Brink, E. N.; Poon, L. L.; Marissen, W. E.; Leung, C. S.; Cox, F.; Cheung, C. Y.; Bakker, A. Q.; Bogaards, J. A.; van Deventer, E.; Preiser, W.; Doerr, H. W.; Chow, V. T.; de Kruif, J.; Peiris, J. S.; Goudsmit, J., Human monoclonal antibody combination against SARS coronavirus: synergy and coverage of escape mutants. *PLoS Med* **2006**, *3* (7), e237.
<https://www.ncbi.nlm.nih.gov/pmc/articles/PMC1483912/pdf/pmed.0030237.pdf>
218. (U) The Novel Coronavirus Pneumonia Emergency Response Epidemiology, T., The Epidemiological Characteristics of an Outbreak of 2019 Novel Coronavirus Diseases (COVID-19) — China, 2020. *China CDC Weekly* **2020**, *2*, 1-10. <http://weekly.chinacdc.cn//article/id/e53946e2-c6c4-41e9-9a9b-fea8db1a8f51>
219. (U) Thevarajan, I.; Nguyen, T. H. O.; Koutsakos, M.; Druce, J.; Caly, L.; van de Sandt, C. E.; Jia, X.; Nicholson, S.; Catton, M.; Cowie, B.; Tong, S. Y. C.; Lewin, S. R.; Kedzierska, K., Breadth of concomitant immune responses prior to patient recovery: a case report of non-severe COVID-19. *Nature Medicine* **2020**. <https://doi.org/10.1038/s41591-020-0819-2>
220. (U) Thomas, P. R.; Karriker, L. A.; Ramirez, A.; Zhang, J.; Ellingson, J. S.; Crawford, K. K.; Bates, J. L.; Hammen, K. J.; Holtkamp, D. J., Evaluation of time and temperature sufficient to inactivate porcine epidemic diarrhea virus in swine feces on metal surfaces. *Journal of Swine Health and Production* **2015**, *23* (2), 84.
221. (U) Thomas, P. R.; Ramirez, A.; Zhang, J.; Ellingson, J. S.; Myers, J. N., Methods for inactivating PEDV in Hog Trailers. *Animal Industry Report* **2015**, *661* (1), 91.
222. (U) Toner, E., *Interim Estimate of the US PPE Needs for COVID-19*; Johns Hopkins Center for Health Security: 2020. <https://www.centerforhealthsecurity.org/resources/COVID-19/PPE/PPE-estimate.pdf>
223. (U) van der Sande, M.; Teunis, P.; Sabel, R., Professional and Home-Made Face Masks Reduce Exposure to Respiratory Infections among the General Population. *Plos One* **2008**, *3* (7). <Go to ISI>://WOS:000264065800020
224. (U) van Doremalen, N.; Bushmaker, T.; Morris, D. H.; Holbrook, M. G.; Gamble, A.; Williamson, B. N.; Tamin, A.; Harcourt, J. L.; Thornburg, N. J.; Gerber, S. I.; Lloyd-Smith, J. O.; de Wit, E.; Munster, V. J., Aerosol and Surface Stability of SARS-CoV-2 as Compared with SARS-CoV-1. *New England Journal of Medicine* **2020**. <https://doi.org/10.1056/NEJMc2004973>
225. (U) van Doremalen, N.; Bushmaker, T.; Munster, V. J., Stability of Middle East respiratory syndrome coronavirus (MERS-CoV) under different environmental conditions. *Euro Surveill* **2013**, *18* (38).
226. (U) Verdict, Cepheid to develop automated molecular test for coronavirus. Verdict Medical Devices: 2020. <https://www.medicaldevice-network.com/news/cepheid-automated-test-coronavirus/>
227. (U) Wan, Y.; Shang, J.; Graham, R.; Baric, R. S.; Li, F., Receptor recognition by novel coronavirus from Wuhan: An analysis based on decade-long structural studies of SARS. *Journal of Virology* **2020**, JVI.00127-20. <https://jvi.asm.org/content/jvi/early/2020/01/23/JVI.00127-20.full.pdf>
228. (U) Wang, D.; Hu, B.; Hu, C.; Zhu, F.; Liu, X.; Zhang, J.; Wang, B.; Xiang, H.; Cheng, Z.; Xiong, Y.; Zhao, Y.; Li, Y.; Wang, X.; Peng, Z., Clinical Characteristics of 138 Hospitalized Patients With 2019 Novel Coronavirus–Infected Pneumonia in Wuhan, China. *JAMA* **2020**.
<https://doi.org/10.1001/jama.2020.1585>
https://jamanetwork.com/journals/jama/articlepdf/2761044/jama_wang_2020_oi_200019.pdf
229. (U) Wang, W.; Xu, Y.; Gao, R.; Lu, R.; Han, K.; Wu, G.; Tan, W., Detection of SARS-CoV-2 in Different Types of Clinical Specimens. *JAMA* **2020**. <https://doi.org/10.1001/jama.2020.3786>
230. (U) Wang, Y.; Liu, Y.; Liu, L.; Wang, X.; Luo, N.; Ling, L., Clinical outcome of 55 asymptomatic cases at the time of hospital admission infected with SARS-Coronavirus-2 in Shenzhen, China. *The Journal of infectious diseases* **2020**.
231. (U) Watson, C.; Cicero, A.; Blumenstock, J.; Fraser, M., *A National Plan to Enable Comprehensive COVID-19 Case Finding and Contact*; Johns Hopkins Center for Health Security: 2020.

- https://www.centerforhealthsecurity.org/our-work/pubs_archive/pubs-pdfs/2020/a-national-plan-to-enable-comprehensive-COVID-19-case-finding-and-contact-tracing-in-the-US.pdf
232. (U) WCS, A Tiger at Bronx Zoo Tests Positive for COVID-19; The Tiger and the Zoo's Other Cats Are Doing Well at This Time. <https://newsroom.wcs.org/News-Releases/articleType/ArticleView/articleId/14010/A-Tiger-at-Bronx-Zoo-Tests-Positive-for-COVID-19-The-Tiger-and-the-Zoos-Other-Cats-Are-Doing-Well-at-This-Time.aspx> (accessed April 6, 2020).
233. (U) Wei, W. E.; Li, Z.; Chiew, C. J.; Yong, S. E.; Toh, M. P.; Lee, V. J., Presymptomatic transmission of SARS-CoV-2 - Singapore, January 23 - March 16, 2020. *Morbidity and Mortality Weekly Report* **2020**, ePub (1 April 2020). <https://www.cdc.gov/mmwr/volumes/69/wr/mm6914e1.htm>
234. (U) Weston, S.; Haupt, R.; Logue, J.; Matthews, K.; Frieman, M. B., FDA approved drugs with broad anti-coronaviral activity inhibit SARS-CoV-2 in vitro. *bioRxiv* **2020**, 2020.03.25.008482. <https://www.biorxiv.org/content/biorxiv/early/2020/03/27/2020.03.25.008482.full.pdf>
235. (U) Wetsman, N., FDA authorizes first antibody-based test for COVID-19. *The Verge* 2 April, 2020.
236. (U) WHO, *COVID-19 Strategy Update*; World Health Organization: 2020. <https://www.who.int/publications-detail/strategic-preparedness-and-response-plan-for-the-new-coronavirus>
237. (U) WHO, Diagnostic detection of Wuhan coronavirus 2019 by real-time RTPCR -Protocol and preliminary evaluation as of Jan 13, 2020. https://www.who.int/docs/default-source/coronaviruse/wuhan-virus-assay-v1991527e5122341d99287a1b17c111902.pdf?sfvrsn=d381fc88_2 (accessed 01/26/2020).
238. (U) WHO, *Infection prevention and control during health care when novel coronavirus (nCoV) infection is suspected*; 2020. [https://www.who.int/publications-detail/infection-prevention-and-control-during-health-care-when-novel-coronavirus-\(ncov\)-infection-is-suspected-20200125](https://www.who.int/publications-detail/infection-prevention-and-control-during-health-care-when-novel-coronavirus-(ncov)-infection-is-suspected-20200125)
239. (U) WHO, Laboratory testing for 2019 novel coronavirus (2019-nCoV) in suspected human cases.
240. (U) WHO, *Modes of transmission of virus causing COVID-19: implications for IPC precaution recommendations*; World Health Organization: 2020. <https://www.who.int/news-room/commentaries/detail/modes-of-transmission-of-virus-causing-covid-19-implications-for-ipc-precaution-recommendations>
241. (U) WHO, Novel Coronavirus (2019-nCoV) Situation Report-5 25 January 2020. https://www.who.int/docs/default-source/coronaviruse/situation-reports/20200125-sitrep-5-2019-ncov.pdf?sfvrsn=429b143d_4.
242. (U) WHO, Novel Coronavirus (2019-nCoV) technical guidance: Laboratory testing for 2019-nCoV in humans. <https://www.who.int/emergencies/diseases/novel-coronavirus-2019/technical-guidance/laboratory-guidance>.
243. (U) Williamson, B. N.; Feldmann, F.; Schwarz, B.; Meade-White, K.; Porter, D. P.; Schulz, J.; Doremalen, N. v.; Leighton, I.; Yinda, C. K.; Pérez-Pérez, L.; Okumura, A.; Lovaglio, J.; Hanley, P. W.; Saturday, G.; Bosio, C. M.; Anzick, S.; Barbian, K.; Cihlar, T.; Martens, C.; Scott, D. P.; Munster, V. J.; Wit, E. d., Clinical benefit of remdesivir in rhesus macaques infected with SARS-CoV-2. *bioRxiv* **2020**, 2020.04.15.043166. <https://www.biorxiv.org/content/biorxiv/early/2020/04/15/2020.04.15.043166.full.pdf>
244. (U) Wölfel, R.; Corman, V. M.; Guggemos, W.; Seilmaier, M.; Zange, S.; Müller, M. A.; Niemeyer, D.; Jones, T. C.; Vollmar, P.; Rothe, C.; Hoelscher, M.; Bleicker, T.; Brünink, S.; Schneider, J.; Ehmann, R.; Zwirgmaier, K.; Drosten, C.; Wendtner, C., Virological assessment of hospitalized patients with COVID-2019. *Nature* **2020**. <https://doi.org/10.1038/s41586-020-2196-x>
245. (U) Wolff, M. H.; Sattar, S. A.; Adegbinrin, O.; Tetro, J., Environmental survival and microbicide inactivation of coronaviruses. In *Coronaviruses with special emphasis on first insights concerning SARS*, Springer: 2005; pp 201-212.

246. (U) Wong, M. C.; Javornik Cregeen, S. J.; Ajami, N. J.; Petrosino, J. F., Evidence of recombination in coronaviruses implicating pangolin origins of nCoV-2019. *bioRxiv* **2020**, 2020.02.07.939207. <https://www.biorxiv.org/content/biorxiv/early/2020/02/13/2020.02.07.939207.full.pdf>
247. (U) Wrapp, D.; Wang, N.; Corbett, K. S.; Goldsmith, J. A.; Hsieh, C.-L.; Abiona, O.; Graham, B. S.; McLellan, J. S., Cryo-EM Structure of the 2019-nCoV Spike in the Prefusion Conformation. *bioRxiv* **2020**, 2020.02.11.944462. <https://www.biorxiv.org/content/biorxiv/early/2020/02/15/2020.02.11.944462.full.pdf>
248. (U) Wu, F.; Wang, A.; Liu, M.; Wang, Q.; Chen, J.; Xia, S.; Ling, Y.; Zhang, Y.; Xun, J.; Lu, L.; Jiang, S.; Lu, H.; Wen, Y.; Huang, J., Neutralizing antibody responses to SARS-CoV-2 in a COVID-19 recovered patient cohort and their implications. *medRxiv* **2020**, 2020.03.30.20047365. <https://www.medrxiv.org/content/medrxiv/early/2020/04/06/2020.03.30.20047365.full.pdf>
249. (U) Wu, J. T.; Leung, K.; Leung, G. M., Nowcasting and forecasting the potential domestic and international spread of the 2019-nCoV outbreak originating in Wuhan, China: a modelling study. *The Lancet* **2020**. [https://www.thelancet.com/journals/lancet/article/PIIS0140-6736\(20\)30260-9/fulltext](https://www.thelancet.com/journals/lancet/article/PIIS0140-6736(20)30260-9/fulltext)
250. (U) Wu, L.-P.; Wang, N.-C.; Chang, Y.-H.; Tian, X.-Y.; Na, D.-Y.; Zhang, L.-Y.; Zheng, L.; Lan, T.; Wang, L.-F.; Liang, G.-D., Duration of antibody responses after severe acute respiratory syndrome. *Emerging infectious diseases* **2007**, *13* (10), 1562. https://www.ncbi.nlm.nih.gov/pmc/articles/PMC2851497/pdf/07-0576_finalD.pdf
251. (U) Wu, P.; Duan, F.; Luo, C.; Liu, Q.; Qu, X.; Liang, L.; Wu, K., Characteristics of Ocular Findings of Patients With Coronavirus Disease 2019 (COVID-19) in Hubei Province, China. *JAMA Ophthalmology* **2020**. <https://doi.org/10.1001/jamaophthalmol.2020.1291>
252. (U) Xinhua, China detects large quantity of novel coronavirus at Wuhan seafood market http://www.xinhuanet.com/english/2020-01/27/c_138735677.htm.
253. (U) Xu, X.; Han, M.; Li, T.; Sun, W.; Wang, D.; Fu, B.; Zhou, Y.; Zheng, X.; Yang, Y.; Li, X.; Zhang, X.; Pan, A.; Wei, H., Effective Treatment of Severe COVID-19 Patients with Tocilizumab. *ChinaXiv* **2020**. <http://chinaxiv.org/abs/202003.00026>
254. (U) Xu, Z.; Shi, L.; Wang, Y.; Zhang, J.; Huang, L.; Zhang, C.; Liu, S.; Zhao, P.; Liu, H.; Zhu, L.; Tai, Y.; Bai, C.; Gao, T.; Song, J.; Xia, P.; Dong, J.; Zhao, J.; Wang, F.-S., Pathological findings of COVID-19 associated with acute respiratory distress syndrome. *The Lancet Respiratory Medicine*. [https://www.thelancet.com/journals/lancet/article/PIIS0140-6736\(20\)30260-9/fulltext](https://www.thelancet.com/journals/lancet/article/PIIS0140-6736(20)30260-9/fulltext)
255. (U) Xue, K. S.; Bloom, J. D., Reconciling disparate estimates of viral genetic diversity during human influenza infections. *Nature Genetics* **2019**, *51* (9), 1298-1301. <https://doi.org/10.1038/s41588-019-0349-3>
256. (U) Yan, C. H.; Faraji, F.; Prajapati, D. P.; Boone, C. E.; DeConde, A. S., In *Association of chemosensory dysfunction and Covid-19 in patients presenting with influenza-like symptoms*, International Forum of Allergy & Rhinology, Wiley Online Library: 2020.
257. (U) Yang, H.; Wang, C.; Poon, L., Novel coronavirus infection and pregnancy. *Ultrasound in Obstetrics & Gynecology* **2020**.
258. (U) Yang, P.; Qi, J.; Zhang, S.; Bi, G.; Wang, X.; Yang, Y.; Sheng, B.; Mao, X., Feasibility of Controlling COVID-19 Outbreaks in the UK by Rolling Interventions. *medRxiv* **2020**, 2020.04.05.20054429. <https://www.medrxiv.org/content/medrxiv/early/2020/04/07/2020.04.05.20054429.full.pdf>
259. (U) Yoon, I.-K.; Kim, J. H., First clinical trial of a MERS coronavirus DNA vaccine. *The Lancet Infectious Diseases* **2019**, *19* (9), 924-925. [https://www.thelancet.com/journals/laninf/article/PIIS1473-3099\(19\)30397-4/fulltext](https://www.thelancet.com/journals/laninf/article/PIIS1473-3099(19)30397-4/fulltext)
260. (U) Yu, W.-B.; Tang, G.-D.; Zhang, L.; Corlett, R. T., Decoding evolution and transmissions of novel pneumonia coronavirus using the whole genomic data. *ChinaXiv* **2020**. <http://www.chinaxiv.org/abs/202002.00033>

261. (U) Zhang, Q.; Zhang, H.; Huang, K.; Yang, Y.; Hui, X.; Gao, J.; He, X.; Li, C.; Gong, W.; Zhang, Y.; Peng, C.; Gao, X.; Chen, H.; Zou, Z.; Shi, Z.; Jin, M., SARS-CoV-2 neutralizing serum antibodies in cats: a serological investigation. *bioRxiv* **2020**, 2020.04.01.021196.
<http://biorxiv.org/content/early/2020/04/03/2020.04.01.021196.abstract>
262. (U) Zhao; Musa; Lin; Ran; Yang; Wang; Lou; Yang; Gao; He; Wang, Estimating the Unreported Number of Novel Coronavirus (2019-nCoV) Cases in China in the First Half of January 2020: A Data-Driven Modelling Analysis of the Early Outbreak. *Journal of Clinical Medicine* **2020**, 9 (2), 388.
263. (U) Zhao, G.; Jiang, Y.; Qiu, H.; Gao, T.; Zeng, Y.; Guo, Y.; Yu, H.; Li, J.; Kou, Z.; Du, L.; Tan, W.; Jiang, S.; Sun, S.; Zhou, Y., Multi-Organ Damage in Human Dipeptidyl Peptidase 4 Transgenic Mice Infected with Middle East Respiratory Syndrome-Coronavirus. *PLoS One* **2015**, 10 (12), e0145561.
<https://www.ncbi.nlm.nih.gov/pmc/articles/PMC4689477/pdf/pone.0145561.pdf>
264. (U) Zhao, J.; Yuan, Q.; Wang, H.; Liu, W.; Liao, X.; Su, Y.; Wang, X.; Yuan, J.; Li, T.; Li, J.; Qian, S.; Hong, C.; Wang, F.; Liu, Y.; Wang, Z.; He, Q.; He, B.; Zhang, T.; Ge, S.; Liu, L.; Zhang, J.; Xia, N.; Zhang, Z., Antibody Responses to SARS-CoV-2 in Patients of Novel Coronavirus Disease 2019. *SSRN* **2020**.
https://papers.ssrn.com/sol3/papers.cfm?abstract_id=3546052#
265. (U) Zhen-Dong, T.; An, T.; Ke-Feng, L.; Peng, L.; Hong-Ling, W.; Jing-Ping, Y.; Yong-Li, Z.; Jian-Bo, Y., Potential Presymptomatic Transmission of SARS-CoV-2, Zhejiang Province, China, 2020. *Emerging Infectious Disease journal* **2020**, 26 (5). https://wwwnc.cdc.gov/eid/article/26/5/20-0198_article
266. (U) Zhongchu, L., The sixth press conference of "Prevention and Control of New Coronavirus Infected Pneumonia". Hubei Provincial Government: 2020.
http://www.hubei.gov.cn/hbfb/xwfbh/202001/t20200128_2015591.shtml
267. (U) Zhou, F.; Yu, T.; Du, R.; Fan, G.; Liu, Y.; Liu, Z.; Xiang, J.; Wang, Y.; Song, B.; Gu, X.; Guan, L.; Wei, Y.; Li, H.; Wu, X.; Xu, J.; Tu, S.; Zhang, Y.; Chen, H.; Cao, B., Clinical course and risk factors for mortality of adult inpatients with COVID-19 in Wuhan, China: a retrospective cohort study. *The Lancet*.
[https://doi.org/10.1016/S0140-6736\(20\)30566-3](https://doi.org/10.1016/S0140-6736(20)30566-3)
268. (U) Zhou, P.; Yang, X.-L.; Wang, X.-G.; Hu, B.; Zhang, L.; Zhang, W.; Si, H.-R.; Zhu, Y.; Li, B.; Huang, C.-L.; Chen, H.-D.; Chen, J.; Luo, Y.; Guo, H.; Jiang, R.-D.; Liu, M.-Q.; Chen, Y.; Shen, X.-R.; Wang, X.; Zheng, X.-S.; Zhao, K.; Chen, Q.-J.; Deng, F.; Liu, L.-L.; Yan, B.; Zhan, F.-X.; Wang, Y.-Y.; Xiao, G.; Shi, Z.-L., Discovery of a novel coronavirus associated with the recent pneumonia outbreak in humans and its potential bat origin. *bioRxiv* **2020**, 2020.01.22.914952.
<https://www.biorxiv.org/content/biorxiv/early/2020/01/23/2020.01.22.914952.1.full.pdf>
269. (U) Zhu, Y.; Chen, Y. Q., On a Statistical Transmission Model in Analysis of the Early Phase of COVID-19 Outbreak. *Statistics in Biosciences* **2020**, 1-17.
<https://www.ncbi.nlm.nih.gov/pmc/articles/PMC7113380/>
270. (U) Zou, L.; Ruan, F.; Huang, M.; Liang, L.; Huang, H.; Hong, Z.; Yu, J.; Kang, M.; Song, Y.; Xia, J.; Guo, Q.; Song, T.; He, J.; Yen, H.-L.; Peiris, M.; Wu, J., SARS-CoV-2 Viral Load in Upper Respiratory Specimens of Infected Patients. *New England Journal of Medicine* **2020**.
<https://www.nejm.org/doi/full/10.1056/NEJMc2001737>